

CEER-A-067

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CEER - A67

CENTER FOR ENERGY AND ENVIRONMENT RESEARCH

NORTH WEST COAST POWER PLANT

QUALITY ASSURANCE MANUAL

February 1980

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February 1980

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?CENTER FOR ENERGY AND ENVIRONMENT RESEARCH

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QUALITY ASSURANCE MANUAL

Sciences Section?

Hi. Ty Be Rosso

REVISION APPROVAL

Toad, Environmental Date quality Assurance Date

Sciences Section

Lead, Environmental Data Quality Assurance Date

Sciences Section

Lead, Environmental Data Quality Assurance Date

Sciences Section

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quality Assurance Menoranda-

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NORTH WEST COAST PROJECT QUALITY ASSURANCE PROGRAM

Organization

The organization of the CEER / North West Coast (SCO) Project group consists of the project office, including the project leader and a small administrative staff, a small logistics action group which handles field operations on shore and at sea, and a number of scientific functional units, each led by a qualified scientist who is specifically trained and experienced in the portion of the

scientific investigation which he directs. Within the scientific

units are some junior scientists and assistants, all of whom have

some training and/or experience in scientific work. Each leader of

an individual scientific unit has responsibility for detailed planning of the investigations that fall within his area of expertise,

for scheduling the appropriate field and laboratory activities and

directing the efforts of his people toward accomplishing the research.

ALL CEER scientific unit leaders are field experienced and routinely

Participate in field work as well as in laboratory work.

The organization of Figure 1 depicts the project organization

and identifies certain key people, including the leader of scientific units.

The Quality Assurance Program will be directed by a Q. A.

Supervisor reporting to the Head, Environmental Sciences Section.

The Q. A. Supervisor will audit the various research activities of

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Center for Energy and Environment Research

Puerto Rico Electric Power Authority North West Coast

(NWCO? Project

Wend, Environmental

Sciences Section

Lt. Tiny

Mead, Marine Biology

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Project Project Manager ?quality

cnier Selentise Rody Zimmerman Assurance

RJ. Zamerman (acting) Ri Rosso,

T

Marine Terrestrial

Ecology Ecology

Thermal / Chemical

1 Quaternary/hydrology

2.3. Zimmerman et al. 3M. Uspex (PD) Fauna

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1 [/ seoente stecies laniton/Fish Studies

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|. Yoshioka (PL)

Buceanse Oven

Garela Garcia

Direction ond Coordination

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(PI) Prineipol tnvesti

Quality Assurance Program Implementation

Organigram For NWCO Project

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the MWCO Project and submit monthly reports to the Director, CHER and

to the Hea, Fnvironmental Sciences Section as outlined in "CEER

Audit Program Plan."

TL. Quality Assurance Program

?The primary function of the CEER Quality Assurance Program will

be to ensure management attention to key elements of the PREPA

Progra for enviroensental investigations for Power Plant siting at

MWCO, The "CER Audit Program Plan" serves as a "Q. A. Manual" for

arch contract. This function will be carried out by means

of a CEER audit program according to an audit program plan which will:

- 1, Identify the individual responsible for the audits
2. Provide for a monthly audit
3. List all activities object to audit and identify specific

ies to be audited in any one month

4, Describe methods to obtain corrective actions on findings

5. Specify documentation and reporting requirements

III. Design control

?The CEER Quality Assurance Program is designed to insure that scientific research work, the schedule for performing the various sections of the work and preparation of reports shall accomplish the

objectives of the PREPA Program for environmental investigations.

?The specific research tasks and the methods to accomplish them are

---Page Break---

outlined in the Specifications for General Performance Requirements

(4) provided by the PREPA.

IV. Control of Documents, Records and Reports

The leader of each scientific unit in the project is responsible for the proper use of the necessary forms, data sheets, log books and other records to fulfill the needs for scheduling and for recording of field trips, samples collected and processed, data taken in the field and taken from field material when processed in the laboratory. The records shall include date, time and location for each sample or measurement. where appropriate, depth will be recorded. Field notes, adequately identified to permit keying to data and

samples, will contain other relevant information such as description

of the site, current conditions such as weather, etc.

The values of the records shall be tailored to the kind of material treated, the type of measurements and methods of sampling and laboratory handling of materials and data. In all cases they shall be such that the leader of the scientific unit can determine upon what raw data all final results are based, i.e. what the raw data values were, and when, where and how they were taken. The raw data values shall be preserved for the life of the contract, and final disposition shall be determined in conjunction with PREPA, ?The leader of such scientific unit is also responsible for controlling the labelling, storage, and handling of his sample

---Page Break---

material from original collection until all data have been extracted

from it and recorded. ?The system used shall be consistent with the rest of his record system and shall permit accomplishment of the stated objectives of identification,

The project leader, assisted by the project office staff, is

Responsible for assembling data and total scientific contributions from all the functional units into the necessary reports to meet the project schedule.

vy

Project instruction

All Leaders of scientific units receive the basic documents which provide guidelines for objectives and methods of the investigation and formats for obtaining and presenting results, e.g. 1

1. Specification for General Performance Requirements for

Aquatic Baseline Studies for Puerto Rico Electric Power

Authority, (PREPAY).

2. Specification for General Performance Requirements for
Terrestrial Ecological Investigation and Analysis for
PREPA.

3+ Specification for General Performance Requirements for
Thermal and Chemical Tolerance Studies for PREPA.

4. Specification for General Performance Requirements for
Tides and Currents Hydrological Data Collection for
PREPA.

---Page Break---

5. North West Coast (SCO) Project Quality Assurance Manual

Items (1), (2), (3) and (4) constitute the prime standard for

the research program:

Copies of reports generated

Led by the project office are distri-

buted to all interested unit leaders.

Contracts

Specified schedule dates for reporting are the accept-

standard for preparation and submission of data

by scientific

units, with allowance for final preparation time in the project office. In addition, the project office keeps unit leaders abreast of dates relevant to their individual functions. Earlier reporting is done whenever data is available.

General operating procedures regarding logistics, communication, safety, etc. are distributed as the need arises by memo from the project office to all unit leaders. Information and discussion meetings for all project scientists are held at appropriate points in

the progress of the work.

Prog:

VI. Control of Equipment

The functional unit leader is responsible for keeping up with the whereabouts and condition of his equipment, giving it proper care and having repairs, servicing and calibration done when necessary.

Where calibration or standardization of measuring equipment can be done satisfactorily by unit personnel, the unit leader insures that such check procedures are accomplished at the proper

6

---Page Break---

finishes and provides the necessary instructions to the people who perform the checks. Where the calibration or standardization procedures is beyond the capabilities of unit personnel and facilities, the unit leader establishes an appropriate schedule based on manufacturer Literature, operating experience, etc. The unit leader is responsible for making the necessary arrangements and insuring that the checks

are made on this schedule or at any other time when a discrepancy

in operation is suspected.

VI, Measuring and Test Equipment Reco

A record of all equipment supplying quantitative data inputs to the BVCO Program will be set-up, These records will include the serial number of each piece of equipment, field and offsite calibration methods, records of calibration, and other pertinent data on the equipment. The unit Leader will keep these records

up-to-date to insure that data

is produced by calibrated equipment.

VIII. Audits

The individual responsible for the audit program will assure that an auditor will review a specified part of the list of activities to be audited, together with additional items such as suspected

weaknesses or follow-up on previous findings. Where possible, the

auditor will not be responsible for work in the area being audited.

The auditor will prepare an audit check list for his own use during

---Page Break---

the audit. In all cases, the auditor will arrange the audit to

minimize disruption of work in progress. An audit will be conduct

ed once each month.

Conducting the Audit - the auditor will use the check list as

a guide during the audit, but will not limit his review to the

specific points listed. For example, in reviewing sample data, the

auditor may find data that is inconsistent; this may lead to questions

on instrumentation. The auditor may obtain information by review

of documents, by discussions with personnel, and by direct observation

of activities. Again, the audit will be conducted to minimize

interference with work in progress. Immediately following the audit,

the auditor will review with cognizant CEER individuals the findings

and recommendations resulting from the audit.

Documentation - each audit by CEER will be reported in a brief,

concise audit report. This report will:

- 1, Identify areas covered by the audit
2. Repore findings in each area
3. Recommend action, if any, to be taken on findings, and
- 4, Report understandings reached during the audit and subsequent review regarding any intended corrective action.

Schedule - while the primary function of the audit program is to ensure proper technical control, audits may identify schedular

problems. Also, constructive audits will define areas where progress

can be improved. Review of audits by senior CER scientists should

include consideration of progress and schedules.

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Scientific peer review - The scientific data generated by the environmental study are subjected to critical review by other

scientists in the appropriate disciplines.

The Q. A. Supervisor will maintain a record of actions agreed upon, the individual responsible for carrying out the changes and the status of corrective action. It is emphasized that the documentation involved in this effort should not be cumbersome, but should enable senior CEER scientists to assess the effectiveness of the program.

TX. Program Records

The following records will be maintained by the CER Quality Assurance Program:

1. Moreh West a

1. Project Quality Assurance Manual and any amendments to it,

2. CEER Audit Program Plan and amendments to it,

3. Schedule of audits of various activities and records of dates when audits were actually accomplished.

4, Completed Audit Check List pertaining to each audit

5. Monthly Audie Reports

6. Internal Correspondence with scientists and supervisors

Yelating to audit program and follow-ups on recoanended

7. Correspondence betwsen PREPA and CEER related to the

Quality Assurance Program

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8. Administrative correspondence related to operation of CEER

QA. Program.

X. Corrective Action

Corrective action will be carried out as stated in the "CEER

Audit Program

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QUALITY ASSURANCE GUIDE-TASK PROCEDURES

SARI ASSURANCE GUIDE-TASK PROCEDURES

The procedures or methods to be used are those presented in

the specifications provided by PREPA, Details for performing various

tasks are included in the following pages.

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Thermal and Chemical Tolerance

1. Checklist of Equipment and Materials

La

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Equipment

111 LAUDA Yodel K-2/R circulating bath units, or suitable equivalents.

1.1.2 ASTM certified mercury thermometers.

78°C to 432°C, total immersions

b. 425°C to 455°C, total immersion

1.1.3, PEARBODY RYAN Model

equivalent recorders,

thermographs, or

1.1.4 YELLOW SPRINGS INSTRUMENTS Model 57 dissolved

oxygen meter, or equivalent, equipped with
YSI 5739 probe, or suitable equivalent.

1.1.5 YELLOW SPRINGS INSTRUMENTS Model 33 salinity
conductivity-temperature meter or equivalent
with YSI 3310 or YSI 3311 probe, or suitable
equivalent.

1.1.6 MASTER FLEX variable speed pump drives, Model
W2IROLS7, or 7364-10, or suitable equivalents,
with pump Models 7013, or 7014, or J015, or
7016, or 7017, or 7018, or suitable equivalents.

METTLER Type KS or K7 top Loading balance, or
suitable equivalent.

2-1,8 MARKSON digital stopwatch - No. 11065, or
suitable equivalent.

Materials

1.2.1 Non-metallic (polyethylene, or
other plastic) collecting and transport containers.

jar) collecting

1.2.2 Aquaria in an assortment of sizes ranging from,
but not limited to, 3.8 liters to 37.8 Liters
in volume.

1.2.3 Metric rules and calipers or equivalent length
measuring devices.

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1.2.4 Flowing seawater, in a climate controlled laboratory. Open or closed circulation provided from

not less than 4,000 liter seawater reservoir, as needed.

1.2.5 Laboratory work tables with drains, designed for flow-through scavenger experiments.

4.2.6 Artificial aeration pumps for aquaria, as needed.

1.2.7 Field and Laboratory notebooks and data entry forms.

2. Important Procedural steps

2.1 Field collection and laboratory holding.

Field collection and laboratory holding.

?Organisms will be collected live in the field using appropriate entrapment gear, species by species (as determined by the project scientist).

?Organisms will be transported to the Laboratory within

the same day as collected in special containers clearly

Ly marked, + Use Seawater Only, No

Small species such as copepods and

be placed in Zip-lock bags (new, double

lined in seawater) in turn placed into larger insulat-

ed containers (not less than 15 liters) with seawater,

~Packing transport organisms will not be excessively crowded or handled, nor will temperature be allowed to rise beyond 1°C above ambient, nor oxygen allowed to deplete to less than 60% of saturation?

?Transfer of organisms to laboratory tanks will take

Place after replacing 252, 502 and 1002 of field

collected seawater with laboratory seawater. In no

case, during transfer will organisms be subject to

more than 3°C temperature change or 3ppt salinity

change in any 12 hour period (EPA specification

See EPA-600/4-78-012, July 1978).

~All organisms will be collected during the week prior to testing. During holding and acclimation, each species will be fed, with food type and quantity determined by a specialist.

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2.2 Acclimation

The pretest acclimation period will not be less than 48 hours, and typically will be 72 hours. If more than 5% of the organisms die within that period, or if they appear to be diseased or otherwise stressed (as determined by the researcher), the entire group will be discarded and a new group obtained (EPA standards; op.cit.).

2.2.1 During acclimation and testing:

2.2.1.1 Live weight loading within each aquarium

will not exceed 2.5 grams per liter.

2.2.1.2 seawater flow rates into aquaria will not be less than one volume exchange each two hour period,

2.2.1.3 dissolved oxygen will not be allowed to become lower than 40% of saturation.

(each of above are EPA standards)

2.2.2 During thermal tolerance testing, three acclimation temperatures are to be used for each species

2.2.2.1 20°C

2.2.2.2 31°C (28°C + 3°C)

2.2.2.3 25°C (28°C - 3°C)

Deviation from these acclimation temperatures

can be instituted if the

extremes are found to be too low or

too high for a particular

species

During chemical tolerance testing,
all acclimations will be at 28°C.

2.3 Tests

2.3.1 Thermal Tolerances

After an acclimation period of 72 hours at
31°C, 28°C or 25°C, organisms are to be placed
abruptly into test temperatures. a

a

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a. 32%

b. 35%

ce. ere

@ are

?+ acclimation control

Deviation from these test temperatures may be allowed

if the scientist in charge deems then inappropriate

for the species being tested

Temperature will be maintained by

Precision heating-cooling, both units equipped with

Variably adjustable thermal regulators positioned in

test aquaria, In addition, the laboratory will be

Temperature controlled to increase precision,

?Test duration will be for 96 hours or until all tests

mortality (excluding the control).

the control organisms die within

the period, the entire test will be abandoned?

?Data will be entered onto prescribed forms similar

to or the same as those appended to this document

designed:

a, mortality data

b. physical test parameters

© equipment check

?These tests are to be considered acute thermal shock

Superinents for the upper incipient thermal Limes

ge distinguished from lower incipient limit and critical

thermal maxima (CTH) experiments. (See: V. #. Hutchison.

Thermal Ecology 1. ERDA Series 40, pages 12 and 1a,

1976).

2.3.2 Chemical Tolerance

?After an acclimation period of 72 hours at 28°C, orga

nisms are to be placed abruptly into 4 test of a Sena]

{eggbe designated) differing by one order of magnitude

(x10) ?each. A separate test without setal ion er

detectable levels will serve as a control.

?Mortality will be recorded on data forms (similar to

those of thermal tolerance teste) over a 96 hours

ais

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period. The test will terminate when, or if, 50 mortality is attained in all test concentration.

The test run will be abandoned if control mortality is beyond 52 during the 96 hours period,

?Tests with three different metals will be run for each species.

?Each metal will be batch mixed week by week at a concentration higher than that of the highest test concentration in @ 4,000 Liter tank. This reservoir will serve as a bioassay test effluent to be minea

in know quantities , using peristaltic pumps into Seawater flowing at?a given rate into test aquaria.

The metallic ion concentration in the effluent reoes= voir will be predetermined by mixing a given quantity

of metal salt, assuming a known dissociation constant,
into seawater. Periodically, samples will be taken
of the seawater in each test concentration and
checked with an Atomic Absorption spectrophotometer,

In general, all procedures for chemical tolerance
testing will adhere, when applicable, to EPA standards
as set forth in Methods for Measuring the Acute

Toxicity of Effluents to Aquatic Organisms (EPA-600/
600/4-78-012, July, 1978)

Report Form

Laboratory data will be entered onto prescribed data

forms similar to, or the same as those appended to
this document. The researcher entering data in each
case will be clearly identified by his or her signature
in a column designated signature or researcher.

and Noe:

Field notes will be entered onto water resistant paper in a standard Engineers Level Book (No. S410V) or suitable equivalent. Each page will be initiated by the researcher taking notes.

All original data will be replicated on a good quality copy machine filed in at least two different places, Pages numbers on each data form will be consecutive and no original data forms regardless of quality will be discarded. If pages are inadvertently lost or destroyed, a full written and signed explanation by the researcher(s) will be inserted in place of,

---Page Break---

4

4.1 Samples

~Organisms collected from the field will be used only once during experiments. Those remaining alive at the end of the 9 hour test will be released into a nearly

habitat similar to that from which they were originally

taken, Dead organisms will be removed from the test aquaria measured as to size and/or weight and disposed of through regular sanitary disposal pickup or into a sink equipped with an ordinary garbage disposal,

~ALL effluent seawater from the flow-through system

Will be directed through an activated charcoal bed

before returning to the environment.

~ALL original data sheets and note books will be deposited with the Environmental licensing Engineer for filing, with copies of data sheets attached to Progress reports to the Uality (PREPA). Copies of ail data and reports as well as comsunications will be kept on file at ckEa.

5. Task Responsibilities of Individuals

3.1 Principal Investigator or Scientist - in charg

(60% effort). Duties included

?design of experiments and decisions regarding modification, if necessary.

?service as specialist on tropical invertebrat:

select organiona for study.

?supervision of Research Associates performing experi-
ments,

?Official communication by direct or indirect means
with all other individuals or organizations:

service as the individuals ultimately responsible for
?IL data collected and research performed,

5.2 Research Associates (two researchers at 100% effort).

Duties included:

IL field collection

?7

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will laboratory culturing and monitoring of organisms,

maintenance and organization of Laboratory according
to accepted scientific standard

?monitoring and data entry for all thermal tolerance
and chemical tolerance experiments

?monitoring and maintenance of equipment calibration

?Preparation statistical analyses of data and Literature:

"Provide consultation and assistance to principal

?investigator in writing the final report.

Service technician

?clean laboratory daily

~saintain ground immediately outside of laboratory

assist in maintaining all non scientific equipment
in the laboratory.

wessist research associates in physical duties,

Maintenance or Calibration of Equipment

6a

Thermal circulation bath units will be checked daily against ASTM Certified Mercury thermometers with deviations noted.

Thermographs will be calibrated initially during a test run with ASTM Certified Mercury thermometers: and checked periodically against the same thermometers to assure repeatable performance.

XSI oxygen meters and probes will be calibrated by the Manufacturer initially. A periodic check on performance will be run using Winkler oxygen titrations (set procedures in Strickland and Parsons, Standard Methods, 1965).

3S? Model 33 Salinity-conductivity-temperature meter will be calibrated against @ standard using TeAve\$.0, Standard seawater and the manufacturer's specification?, and checked weekly.

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6.5

6.6

MASTERFLEX variable speed drives and peristaltic pumps

Will be checked daily, and flow rates determined using

PYREX graduated cylinders calibrated by the manufacturer

in milliliters, and a digital stopwatch (manufacturer

calibrated) measuring in not less than 0.1 second

intervals.

METTLER top loading balance to be calibrated prior to

use by a certified manufacturer's technician.

Unless otherwise specified, maintenance and calibration

of equipment will follow manufacturer's specifications,

Hi

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APPENDI: BIOASSAY DATA FORM

?Thenaa/Chemical Studies Marine Ecology Division

FORMAT T ceer, UR

Mayeguer, #8

TEST ORGANISH

INITIAL NUMBER OF TWOIVIDUALS

LIFE stage

SIZES (3, #50, range)

I

DATES: INITIAL

ACCLIMATION TEMP,

TEST TEMP,

EFFLUENT,

____. comeere

+ ONC,

i

TEST 10,

_ = REPLICATE No. .

oo ooo

oe | TIME | ALIVE | worraLtry ?OBSERVATION RESEARCHER

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TEST W0..

TEST PARAMETERS;

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APPENDIX

THERMAL CHEMICAL STUDIES

FORMAT TT

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+ REP. HO.

SIGNATURE RESEARCHER

con.)

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20

36

48

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---Page Break---

IT, Benthic Studies

1. Checklist of Equipment and Materials

11 Equipment

haa

tae

Ls

inet

Ls

SCUBA gear: tanks, regulators, accessories

Underwater communication system: Wet phone

Sound wave System, Inc.

Underwater camera: Nikonos Model 111

Underwater strobe: Model Sunpack marine

Microscopes

1.1.5.1 Compound: American Optical: Series

H10 Microstar Model X¥10 TM = JW.

with Canon:

Attachment 1053, 35 mm with magnifi

cation factors of 2.8 x and 5.0 x. With

exposure meter series 1056, Expostar

Control units (115-120v, 6 Okg, series

1190,

1.1.5.2 Dissecting: Sausch and Lomb: Sterozoom

7, from Baush and Loab Tne.

12 Maceriats

had

1.2.2

123

Cement blocks

Asbestos panels

Chains

Buoys

Chesicals

Containers,

2, Important Procedural seeps

?The specifications established by the engineer for
the study of the benthic flora and fauan at the Rincdn and

2.

---Page Break---

Carrigal sites are stated in pages 23-28 of 85. There are three major specifications for both sites: A. benthic Mappings B. Sampling Stations and C. artificial Substrates.

The methods to meet these requirements are given below.

2.1 BENTHIC HAP: The methods to create a benthic map for both sites are given below.

21.1 Aerial Photography: Aerial photographs will be used for (a) general reconnaissance of each site;

- (b) determining general shoreline features
- (c) determining the approximate location of submerged reef structures; submerged rock platforms and rock outcrops in relation to clear (i.e. sandy) bottoms, Both black and white film (Plus X, ASA 125) and color film (Kodachrome, ASA 64) will be used for this purpose. PREPA's helicopter or an airplane (preferably from West Wing Flying School, since it is close to both sites) is required,

2.1.2 Bottom Profiles: bottom profiles will be determined aboard the R/V SULTANA. The instrument, already installed on board is a Ross Fisherman Depth Finder and Recorder. The advantage of this instrument is that, in addition, it will provide information on substrate type since it can distinguish between soft and hard bottom:

A soft or silty bottom will appear on the chart, as a relatively narrow bottom line due to absorption of the acoustic signal, a hard or rocky bottom will have a much wider line at the same sensitivity setting due to higher bottom reflectivity of the acoustic signal.

2.1.3 Underwater Transects: Underwater transects are required by the engineer at both sites (pages 23-25 of 85) "to evaluate substrate type, biotic associations, and zonation" (page 25 of 85). These transects will be useful for creating benthic maps at both sites. The consultant

proposes the following methodology to comply with these specifications. The location and direction of each transect is given in Fig. 2-1-1 (Rinedn) and in Fig. 2.1.2 (Carraizal). ?Transect Lines Will be 300 m'long marked at 20m intervals. They will run from shore to offshore.? A format (FORT 1, Appended) has been prepared to meet

2

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FORMAT 1

Dates

Bearings end of Te: Station:

Depth Rang. ?Type of Sample:

Signature:

Conspicuous

Distance(s) Depeh(fe) _ Substrate Species

om

0-2

20 - 40

40 - 60

60 - 80

80 - 100

100 = 120

120 - 140

140 ~ 160

160 - 180

180 - 200

200 ~ 220

220 - 240

240 - 260

260 ~ 280

280 ~ 300

300m

---Page Break---

FORMAT 11

Bearings of Station: Date:

Depth Range:

Distance from Shore:

Signature:

Conspicuous

Depth Substrate DoM-coves Species

25

---Page Break---

2.2

the specifications mentioned above. U/W photo
graphy Will also be used to characterize the
bottom. Samples of conspicuous, unrecognizable

organisms will be collected.

2.1.4 Spot Dives: As many spot dives (hereby defined as dives at preselected spots) as seen

Necessary to fulfill the information required

to construct a benthic map will be made. A

format (FORMAT II) has been prepared to document,,

underwater, the specified information. Underwater

photography as well as samples of conspicuous,

unrecognizable species, will be taken.

Sampling Stations: Sampling stations on predetermined

locations have been selected by the engineer to provide

information on more specific aspects of benthic community

structure and dynamics: a) species present (invertebrate,

algae, fish; b) species interaction of major species

Present; c) ?seasonal changes in biotic components;

4) changes in the biotic components and 3) population

densities of conspicuous species (pages 26-27 of 85),

In order to meet these requirements, the engineer

Proposes a 10 m x 2 m transect as the major sampling

area, The consultant proposes the following methodology

to meet the requirements of the engineer? utilizing the

10 m x 2 m transect proposed.

Three 10 m long chains spaced 1 m apart from the central

chain, each running parallel to each other, will be

Permanently secured to the substrate 90° to provide

the 10 m x 2 m area specified by the engineer in page

26 of 85. At every meter a transverse line will be

placed, and secured, so as to provide twenty 1 m² plots

in two rows: ROW I and ROW II, each consisting of 10,

1m² plots. Each plot is enumerated as illustrated

in Figure 1 (appended). At each sampling station ROW

will be photographed twice yearly; at the beginning of the study and 12 months after the beginning of the study, and the area described in detail so as to note any changes in biotic components as specified in page 26 of 85. Random collections during the year will be obtained. Ten, 1/16 m², quadrats will be taken at

the beginning of the study and 12 months after the beginning of the study. Counts (population density)

of conspicuous plants and animals will be determined within the 10m x 2m transect, at the beginning of the study, and 12 months after the beginning of the study,

FIGURE 1

19

20)

2

---Page Break---

2.3. ARTIFICIAL SUBSTRATES: In section 2.2.1.4 pages 27-28

of 85 the engineer specifies the requirement of an artificial substrate study in order to give an idea Of the potential for recolonization at predetermined Stations. The Duratex panels will be tied to the surface of the cement blocks and will cover the same surface area as the surface of each block. The blocks will be aligned in two rows: one of 7 blocks and another row of 6 blocks. A chain will run through the holes of the blocks through the whole length of the row and secured to the substrate. The sampling procedure has already been established by the engineer (pages 27-28 of 85), The Rine6n site is a high wave energy environment, and modifications of securing the blocks to the substrate may be necessary.

Report Forms and notes

Report forms and notes have already been specified in

"specification for General Performance Requirements for Aquatic Ecology and Water Quality: Aquatic Baseline Studies? Section 6, pages 61-85 of 85, Revision 2. The report forms are given in Fig. 6.1.1 and 6.3.1 and the species code assignments in Table 6.3.2

Disposition of Samples and Results

As specified in Section 2.2.24: Benthic Flora and Fauna (Contractor's Specification), all specimens collected in the field will be taken to the laboratory and identified to Species. A list of all reference used and systematists consulted will be reported to the owner and the engineer by the consultant. All species data shall be recorded on the Biological Analysis Report forms provided by the environmental Licensing engineer (Appended).. Specimens collected will be kept in the laboratory, preserved in the proper agents (Zornalin, ethanol) and labeled for station number and date, A reference collection for each group will be set up.

?Task Responsibility of Individuals

5.1 Principal Investigator. In charge of designing field studies, supervision of all aspects concerning the benthic studies; laboratory responsibilities also include taxonomy of the Porifera; submission of monthly

reports.

---Page Break---

5.2

5.3

Maintes

Scientific Research Associate III. 19 charge of all invertebrate taxonoay and coordinator of laboratory duties (proper labeling of specimens, proper allocation of samples).

Lab. Tech IL.

actophytes|

In charge of the taxonomy of all

Field assistant when necessary.

Lab. Tech. T. In charge of obtaining and preparing
all field materials. Principal field assistant in all
Phases of SCUBA diving activities.

\ce and Cali

Maintenance and calibration of equipment will follow

manufacturers specifications

29

---Page Break---

[APPENDIX: BENTHIC STUDIES

inited e:

BIOLCSICAL FIELD COLLECTION REPORT FOP

ve pero SSeS Mee erm

souneres Specification No, 6212-000.2-8-5-F

Tape 62 of 85

Revision 2

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xX: eNTATE sTUDIES

hited engineers

Figure 6.41 BIOLOGICAL ANALYSIS REPORT FORM

eee

oe kevicion ?2

---Page Break---

APPENDIX: BENTUIC STUDTES

TABLE 6.3.2

SPECIES CODE ASStoMMMENTS

Loses yp

Tae

() The first space will tell the computer what general group the species belongs to:

ot used

+ Rot used (signifies water quality)

Phytoplankton,

Zooplankton

1

2

3.

4

5. Tehthyep plankton

6. Benthic Fauna

7. Fish

8. Benthic Flora

(2,3,4) Spaces 2, 3 and 4 are used to designate the genus order for

each particular group

(5, 6) Spaces 5 and 6 are used to di

genus.

ignate species within each

Specification No. 6112-020-2-8-5-F

---Page Break---

MIT. Water quality / Hydrology

1, CheckList of Equipment and Materials

Lal Equipment

Maa

1120

hin

Let

hrs

add

Laas

Laae

Lay

Secchi Disk, standard 20 cm diameter oceanographic type

YSE Model 57 Dissolved oxygen meter or equivalent

DRT-200 Turbidimeter or equivalent

Corning pH meter or equivalent

YSI Model 33 Salinity-conductivity-Temperature meter or equivalent

Millipore Bacteriological Analysis kit or equivalent

Precision Scientific Freas 815 Tacubator or equivalent

Mettler H 542 Analytical balance or equivalent

Kontes MicroKjeldahl digestion system or equivalent

?Technicon Auto Analyzer IT or equivalent

Perkin Elmer Model 303 Atomic absorption

Spectrophotometer and accessories or equivalent

Coleman/Perkin Elmer MAS 50 Mercury Analyzer

or equivalent

Niskin Water Bottles 51 capacity - non metallic

or equivalent.

Millipore vacuum filtration pump or equivalent

Soxhet Extraction System for COD or equivalent

Beckman Model 3600 U/VIS Spectrophotometer or

equivalent

Millipore Milli RO4 water purification or

equivalent

33

---Page Break---

12 Materials

1.2.1 Polyethylene sample storage bottles, cubitainers

1.2.2 30D bottles numbered (glass)

1.2.3 Glass sample storage bottles for coD

1.2.4 Millipore filter holders and 0.45 um pore size
membrane filters or equivalent

1.2.5 Burettes, beakers, volumetric flasks and other
glassware

1.2.6 AA Standard Solutions for Trace Metals (Fisher
certified or equivalent)

1.2.7, Analytical Grade reagents as required in Standard
Methods for all analyses.

2. Important Procedural steps

2.1 Field Collections-Rincon: Water Samples

Water samples for analysis of water quality

Parameters, will be collected sonthly at Station 0230

as described in the Licensing engineer's specitcat ions

surface (nominal depth of 0.5 a) and near the bottom

(8.0 m) depth at Station 0230 as in specification

Replicate samples will be collected in sequential

ts. A nonmetallic water sampler such as a Poly vinyl chloride Miskin type sampler will be utilized. The capacity of the sampler will be of sufficient volume to collect all necessary sample aliquots in a single cast. The capacity of the sampler will be large enough to allow collection of a 500 ml sample aliquot for Phytoplankton enumeration and plant pigment samples at Station 0230 (as in specifications) surface.

Water samples will be collected after three rinses of the sample bottles previously acid rinsed and distilled water rinsed. All water samples will be stored in a freezer on board the sampling vessel, and frozen until analysis as a means of preservation.

ALL sample containers will be clearly labeled with location, code, station number, depth of collection, time, date and collector's initials.

a3

---Page Break---

2.2

2.3

Profiles and Field Measurements - The researcher will measure temperature and dissolved oxygen with a YSI Model 57 Dissolved Oxygen Meter, or an equivalent, at the surface (nominal depth 1.0 feet) and at 1-0.5 meter intervals to the bottom at Station 0230 (as in specifications) on a monthly basis. The researcher will measure Secchi Disk Transparency, Turbidity and pH at Station 0230 on a monthly basis. Profiles and Field measurements will be made concurrently with water sample collections.

Field Collection ~ carrizal

Water samples - ater sanples for analysis of water quality parameters, shall be collected monthly at Stations 0130 and 0230 (as in specifications). In addition, at Least, one collection for all physical and chenical paranctere (described in specifications) will be made during a period in which Rio Culebrinas is in a flood condition. Collection will be made at the surface (nominal depth of 0.5 m) ané near the bottom (8.0 m depeh) at both Stations 0130 and 0230 (as in specifications). Methods and type of sampler will be the same for Rincon, however the capacity of the sampler will be Large enough to collect water quality and phytoplankton samples ar Station 0230 surface and bottom.

Profiles and Field Measurements - Measurements|

Will be made at both Stations 0130 and 0230 (as describ=
ed in specifications).

Analytical Methods

State of the art analytical techniques will be used. Water Quality parameters will be determined by methods prescribed in

APHA, AUWA, WPCF ?Standard Methods for the Examination of Water and Wastewater? ashington, D.c, 1976,

Strickland, J.D.H. and T.R. Parsons. "A Practical Handbook of Seawater Analysis" Bulletin 167 Fisheries Research Board of Canada, Ottawa (1968).

US EPA "Manual of Methods for Chemical Analysis of Water and Wastes? Office of Technology Transfer, Washington, 1972,

235

---Page Break---

3. Report Forms and Wot

3:1 Field Notes - Field notes will be taken in conjunction
With the water quality sampling. Water bottle sampling
notes and Secci Disk Transparency Turbidity and pit will
bbe recorded in No. 2 pencil on water proof field. sheet
the following information at each water quality sampling
Station: date, time of day (local 24-hour time) tidal

ge from USCOS tables, wind speed and direction, wave
height and direction and weather observations. Notes

WAIL be made of any unusual occurrences which might
affect the quality of the samples collected, Temperature

and dissolved oxygen profile data will be recorded in a
water proof field notebook. Each log sheet and field
notebook page will be signed or initiated by the
individual responsible for the collection and handling
of samples in the field.

4, Disposition of Samples and Reports

Water samples are kept frozen until analyses are performed and then are disposed of.

ALL original data sheets and notebooks will be deposited with the Environmental Licensing Engineer for filing, with copies of the data sheets attached to progress reports to the owner. Copies of all data and reports as well as communications will be kept on file at CEER.

5. Task Responsibilities

5.1 Principal Investigator or Scientist in charge ~ Duties:

= Responsible for supervision of research associates performing sampling and analysis

= Serve as specialist in analytical, marine and environmental chemistry.

= Design and/or modification of sampling or analytical methods

= Official communication to all other individuals of organizations.

5.2 Research Associate

= Maintenance and organization of the laboratory

---Page Break---

+ Instrumental and wet chemical analyses

~ Statistical analysis of data

> Assist principal investigator in report writing

= Field sample collection

5.3 Laboratory Technician

~ Assist research associate and principal investigator

in all aspects of field sampling measurements and laboratory analyses.

Maintenance and Calibration of Equipment

All calibrations to be performed on equipment will be in accordance with the manufacturer's specifications or

using materials and methods that are traceable to the National Bureau of Standards and state of the art techniques,

37

---Page Break---

IW. Plankton / Fish studies

A, Zooplankton and Ichthyoplankton

1. Checklist of Equipment and Materials

1.1 Field collections

haa

hua

1a

hae

has

Net frames, nets, flometers

borties

labels

data sheets

stop watch

formalin

hose for washing nets

wateh

1.2 Laboratory

Lat

122

123

128

1.2.5

1.2.6

2.1 Field

2a

21

microscopes

counting tray

subsamples (Folson splitter & Stempel

pipette)

bottles

data sheets

Important Procedural steps

Nets will be rigged according to manufacturer's specification:

Flowmeter will be read prior to launching of net.

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---Page Break---

2.1.3 time, depth, seastate will be noted on data sheets,

2.1.4 nets will be lowered until proper depth is reached and will then be opened.

nets will be towed until a minimum of 100m?
of water is filtered.

2.1.6 nets will be closed and raised to surface

2.1.7 flowmeter will be read and time recorded

2.1.8 nets will be rinsed

2.1.9 sample will be placed in sample bottt
containing labels and formalin

2.2 Laboratory

2.2.1 pi of samples will be checked as soon as
possible after their return from the field

Aliquots will be taken with the Stempel
Folsom plankton splitter or pipette,
depending upon the size of subsample requir-
e to count a sufficient number of plankton
(= 300).

2.2.3, Identifications will be Listed on the data
sheets and signed by the identifier.

Report forms and notes

3-1 Report forms and notes will be kept following the
Procedure outlined by the engineer.

Disposition of samples and reports

4.1 Zooplankton samples will be kept in a collection

4.2 Reports will be made available to the chief
scientist to be relayed to the owner or engineer

Responsibility

5.1 Principal Investigator - administrative tasks, budget management, and overall scientific supervision of program

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---Page Break---

5.2. Scientist - supervise collection and processing of samples. Design of specific scientific studies
Analysis of data

5.3 Laboratory techniques - process samples and record data. Cataloguing of samples. May aid in field collection.

5.4 Field technician - aid in field collection, records some field notes, maintenance of equipment. Aids in equipment calibration.

6. Maintenance and Calibration

Maintenance and calibration of equipment will

Follow manufacturers specification.

3. ian

2. Checklist of Equipment and Materials

La Flee

LiL Goh erape with Lines and floats

1.1.2 diving equipment ~ tanks, regulators,

Backpacks masks, and fins ~ then transecte

11.3 data sheets

1st.6 plastic tags and dabets

1.2. Laboratory

Litt atance

1.2.2 moter stick

1.2.3 data sheets

1.2.4 Soteies

1.2.5 tommatin

2. Important Procedural Steps

2.1 Fiead

toe

---Page Break---

2.1.1 Fish traps to be placed at sites designated
by engineer

2.1.2 fish traps will be in place overnight or for
24 hours conditions persitt ing

3. Report Forms and Notes

Report forms and notes will be kept according to
directions of the engineer, and will be available ts
the owner of engineer.

4. Disposition of Samples and Reports

Samples will. be discarded following the foraation
of a feference collection. Reports vill be given to the
chief scientist to be forwarded to the omer or engineers

5. Task Responsibility

5-1 Principal Investigator - Administration and budget

Management Scientific supervision of program

5.2 Research Associate ~ carries out program under supervision of PI. Carries out sampling program, identifies and weights fish. Fills out data sheets. Supervises field technician.

5.3 Field technician ~ Aids Research Associate, Maintains equipment. Records some field notes:

Maintenance and Calibration

Maintenance and calibration of equipment will follow manufacturers specifications:

©. Phytoplankton

1 Checklist of equipment and materials

1a Field

11.1 Miskin bottles

1.1.2 containers

1.1.3 formalin

mle

---Page Break---

1.1.4 vacua pumps and filters

11.5 Mgc0s

1.1.6 envelopes

L.L.7 freezer, refrigerator, or ice chest

1.1.8 Labels, field data sheets

2.1 Laboratory

2.1.1 Inverted microscope

2.1.2 settling chambers

2.1.3 fluorometer

2.1.4 acetone

2.1.5 data sheets

2. Important Procedural Steps

2.1 Utermohl procedure will be used to count phytoplankton.

2.2 Fluorometer will be used to measure chlorophyll and phaeophyta following Strickland and Parsons.

3. Report Form

Report forms and notes will be kept following

Procedures outlined by the engineer.

Disposition of samples

Samples will be discarded due to their degradation

with time, Results will be relayed to the Chief

Notes

Scientist.

5. Task responsibility

5.1 Principal Investigator ~ supervise project,
administration and budget management

5.2 Phytoplankton taxonomist - count and enumerate
phytoplankton

more

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5.3 laboratory technician ~ measure chlorophyll and

Phaeocystis concentrations on the fluorometer

Maintenance and calibration of

Maintenance and calibration ϕ

Fluorometer will be calibrated against @ spectro=
Photoneter following manufacturers specifications.

43

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Terrestrial Ecology ~ Vegetation and Fauna

1, Checklist of Equipment and Materials

La

Le

Equipment

1.1.1 Tomshawk collapsible Live traps (6 x 6 x 24 inches)

1.1.2 Sherman Live traps (large)

1.1.3. Victor snap traps

Ld Mist nete

1.1.5. binoculars

1.1.6 portable tape recorder

11.7 35 om camera

11.8 headlight

1.1.9 quadrat frames

1.1.10 calipers

L111 insect light traps

1112 sticky traps

2.113 Tullgren extraction apparatus

1.1.16 plane press

Materials

1.2.1 bait (Peanut butter, eggs, bird seed, etc.)

1.2.2 Fite

1.2.3 drafting supplies

1.2.4 tapes

1.2.5, herbarium materiale

1.2.6 preservatives (formalin, alcohol)

1.2.7 drift fence and pit trap materiats

he

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2, Important Procedural steps

The specifications established by the engineer for the study of terrestrial flora and fauna at the Rincon and Carrizal sites are stated on pages 5, 5A, and 6 of the Specifications (Revision 2) for Terrestrial Ecology Investigation and Analysis. Table 1 presents the sanpling and report shcedule,

The methods which CEER's Terrestrial Ecology Division will employ to meet these specifications are given below:

2.1 Vegetation Map

21.1 Aerial photography (supplied by the owner) will be used for a general reconnaissance of each site, the ash disposal pile, and proposed rights-of-way. Additional photographs will be taken of the sites to document and map recent changes, PREPA's helicopter or airplane (preferably from West Wing Flying School) is required for 2-4 hours of overflight time. All photographs will be used to construct a map which indicates the distribution of vegetation types (including cropland), major topographic features, and area of disturbance,

Ground-truthing surveys will be conducted to field check a draft map and identify problem features seen on aerial photos. These surveys will be conducted in conjunction with vegetation inventory surveys,

2.2 Biological Inventories

The major seasonal periods mentioned in the Specifications are hereby defined as Wet season (July-September) and Dry season (January-March). Intermediate seasons are transitional between these and are defined as minor. Unless otherwise specified, all sampling locations will be randomly located using a grid (Phillips, 1959),

2.2.1 Vascular Plants - Limited sampling will be conducted during major and minor seasons (total of four Surveys) to collect specimens and compile a detailed species list for each area of proposed development.

Vegetation types on each site and the ash disposal Pile will be quantitatively sampled during the two

mis

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major seasons. A species-area curve will be constructed for each vegetation type to ensure that the species composition is adequately represented. The sample size will be considered adequate when an increase in 10% of the total area yields only 10% (or fewer) more species (Cain, 1938; Mueller-Dombois and Ellenberg, 1974).

Woody vegetation (trees and shrubs) will be sampled using the point-quarter method (Cottam et al., 1953). A

minimum of five 50m transects (10 points each) will be sampled in each major season in each vegetation type dominated by woody species. Woody vegetation in other types will be mapped and total counts will be made.

The species, point-to-plant distance, basal area (for trees), and crown diameter (for shrubs) will be recorded and used to calculate relative density, relative dominance,

relative frequency, density, and total basal area

(Phillips, 1959).

Herbaceous vegetation will be sampled using a minigun of twenty 1 m quadrats in each vegetation type. Data on cover, frequency, and density of species will be recorded and subsequently calculated according to standard methods (Cox, 1967).

Voucher specimens of plants which cannot be identified in the field will be collected and preserved according

to standard procedures. These specimens will subsequently be identified by recognized experts.

2.2.2 Faunal Inventories

Field sampling will be conducted during each of four seasons. Intensive sampling including trapping will be conducted during major seasons only and will be confined to the two proposed sites and associated ash disposal pile. All major habitats will be surveyed. Faunal habitats include all major vegetation types plus specialized areas

Such as wetlands, rocky outcrops, and caves. Less extensive surveys for most groups will be conducted quarterly to collect additional information on the presence and seasonal use of habitats by faunal species. Random observations of animals seen

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within study areas will be recorded and used to augment standard sampling data.

Unless otherwise specified, all sampling locations will be randomly selected within each habitat type. Voucher specimens of selected animal species (except birds) will be collected and preserved according to standard museum procedures (Hall, 1962; Jacques, 1947). Species identifications will be made in the field whenever possible. Unusual individuals and specimens which

ate difficult to identify will be collected for subsequent verification by recognized experts. Records of threatened and endangered species occurrence will be documented by photographs and verified by experts to the extent practicable.

2.2.2.1 Mammals

One or more traplines will be established in each major habitat type. Each

line will consist of 10 live traps placed in a straight line at intervals

of approximately 15m. Traplines will

be checked each morning for three or

four consecutive days during each of the two intensive sampling periods. Captured individuals will be marked and released, and the traps will be rebaited at each trap check. The date of capture, species, location and habitat affinity of each

individual will be recorded (Figure 1).

Minor habitats and habitats situated along
Rights-of-way (ROWs) will be sampled using
selective Live and snap trapping procedure
Live captured individuals will be marked
and released.

Bats will be sampled by mist netting at
selected locations within study area.

To the extent feasible, some individuals
may be collected by shooting. Potential
roosting areas (e.g. caves and abandoned
buildings) will be visited to determine
species presence and seasonal «

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2.2.2.2

2.2.2.3

Birds

Standard methods (Franzreb, 1977) will be used to gather information on the presence, habitat affinities, seasonal use, and general abundance of bird species.

Replicate surveys will be conducted within each major habitat type during periods of breeding, migration and overwintering.

Resident breeding species and seasonal migrants will be identified. Important avian habitats (e.g. wetlands) within one kilometer of designated study areas will be also be sampled.

Qualitative surveys will be conducted quarterly within each study area so that

@ more complete species list can be compiled. Survey methods may include

walking surveys, the use of recorded calls,
and limited mist netting. Sampling
Procedures may be modified to conform to
existing conditions in each study area

Amphibians and Reptiles

Diurnal species such as anoles will be
surveyed during the dry and rainy seasons
along randomly located transects in each
major habitat type. Intensive searches at
selected locations within each study area
will also be conducted to detect the pre-
sence and habitat affinities of secretive
and fossorial forms. Surface debris will
be established in selected locations to
sample ground dwelling forms. Limited
collecting may be required in order to
verify species identifications.

Each study area will be visited at night
during the rainy season to detect calling

anurans (frog and toads). Most species identifications will be made in the field by trained observers. Calls not readily identifiable in the field will be recorded for subsequent expert verification. A few voucher specimens of amphibians and reptiles may be collected and preserved.

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2.2.2.4 Invertebrates

The sampling program for terrestrial invertebrates will concentrate on insects. Field surveys will be conducted quarterly. Light traps and sticky traps will be used to collect representative samples from each of the designated study areas. These techniques will be supplemented with netting, vacuum sampling, and surface collecting during the two major Sampling periods.

Soils and leaf Litter arthropod sample:
will be collected during the two intensive
sampling surveys, Arthropods will be
Yenoved from samples using the funnel
extraction techniques (Cox, 1967)
Extracted organisms will be identified and
counted.

Records will be kept on the habitat
affinities ané general abundance by major
groups (e.g. families, important generals
ete.) for each season. Insect collections
Will be examined by project staff, and
identifications subsequently verified by
recognized experts.

2.3 Important Species

Lists of candidate important plant and animal
species potentially inhabiting areas of proposed project
development will be submitted within days after the

first field visit. The List will be based on a review of published Literature, discussion with regional authorities, and an observations made during a reconnaissance survey of the study areas. The List will be updated following each field survey.

A species will be considered important if it ia:

2.3.1 Commercially or recreationally valuable,

2.3.2 officially designated

threatened or endangered,

2.3.3 Likely to affect the vell being of an important species as defined in 1 or 2 above, oF

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2.3.4 sufficiently sensitive to induced environmental

stress that it could serve as a stress indicator

before significant effects on other important

species occur.

Species which are thus defined will be emphasized during baseline characterization studies. If initial surveys indicate that project development may significantly affect the well-being of one or more important species CEER would propose additional quantitative studies beyond the present workscope in order to determine the magnitude of the predicted impact,

Report Forms and Notes

ALL field report forms and notes will be dated and signed by the personnel responsible for collecting the data. Copies

of all field notes will be maintained in a central file and originals handled in the manner indicated in the Specifications document. All field notes and data sheets will be initialed by the appropriate principal investigator to indicate that

they have been approved.

Disposition of Samples and Results

ALL specimens will be properly preserved (e.g. plants on herbarium sheets, animal specimens in alcohol, formalin, or Prepared as study skins) according to the museum techniques referenced in the appropriate methods sections. All voucher specimens and results will be maintained at CEER facilities in Mayaguez, Rio Piedras, or El Verde and supplied to the owner upon request.

?Task Responsibilities of Individuals

5-1 The two principal investigator (PIs) will be Douglas Reagan (fauna) and Susan Silander (vegetation). Each PI will be responsible for conducting and supervising all aspects of their respective tasks.

5.2 Task leaders will be in charge of data collection, Specimen preservation, and data analysis for their disciplines. Task leaders are: Susan Silander (vegetation studies), Robert Waide (birds), Douglas Reagan (mammals, reptiles, and amphibians), and Miguel Canals (invertebrates).

5.3 Technician level field assistance will be required, but individuals are not specified, Responsibilities will be to assist the task leaders in the conduct of the various tasks

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Maintenance and Calibration:

ALL equipment will be maintained according to manufacturers' specifications:

properly maintained according to manufacturer

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TABLET

TERRESTRIAL ECOLOGY SCHEDULE

00

Task / Report Date

Reconnaissance Survey March 1-31, 1980

Submit Candidate Important Species List within 15 days of

Spring Field Sampling (minor)? april 15 ~ June 15, 1980

Wet Season Field Sampling (major)? July 15 ~ sept. 15, 1980

Fall Field Sampling (minor)? Oct. 15 = dec. 15, 1980

Submit Site Characterization Report by November 30, 1980

Dry Season Field Sampling (major)? January 15 ~ Feb. 15, 1981

Final Data Report by March 31, 1981

Ba

1, Assumes contract issued March 1, 1980; 13 months required to complete project tasks

2. A seasonal biological inventory report will be submitted to the engineer within 30 days? of completion of all field work.

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Ww.

Proviso for Procedural Changes

Proviso for Procedural Changes,

Procedural changes that will affect results or data procured

can be made with the approval of the Omer and Engineer.

Minor changes in procedures that will not affect the results

OF data outcome can be made at the discretion of the scientist~

in-charge or of the principal inv

tigator.

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CER AUDIT PROGRAM PLAN.

?The Audit Progran Plan is designed to fulfill the requirements of the PREPA Quality Assurance Program for the Norch West Coast ({WiCO) Project. The position responsible for the CEER Audit Program will be the Quality Assurance Supervisor who will report directly to Head, Environmental Sciences Section,

(One of the activities listed below will be audited each month.

The report of the Audit, with recomendations for further action,

and follow up on previous recommendations will be available to the SWCO Project Manager and to other concerned scientists within three

(3) weeks from the date of the audit.

To The 5

scientific activities (scientific units) to be audited are:

Terrestrial Ecology ~ Vegetation and Fauna

Plankton / Fish Studies

3. Water Quality / Hydrology

?Thermal / Chemical

Benthic Studies

?The activities to be audited each month are as follows: arch-
#L, April - 42, May - 65, June ~ % and July - #5. The schedule will

repeat this sequence unless experience shows the ns

for a change,

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When an Audit shows that corrective action is nect

ry, the

Proposed corrective action will be discussed jointly with the leader
of the scientific unit and the MiCO Project Manager hen @
course of corrective action is agreed upon, it will be summarized
in writing with copies for each person involved. If @ course of
corrective action cannot be agreed upon, the Head, Environmental
Sciences Section will join the discussion and decide on what action
Af any is necessary. It is the responsibility of the Quality
Assurance Supervisor to follow up in any corrective action to

determine if it is accomplishing its objective

Whenever an Audit results in corrective action in relation to the work of a scientific unit or other function of the MHCOP Project, this corrective action will be followed up within two months either as part of a monthly Audit or directly. The extent of application of the corrective action and its effect will be included in the next Monthly Audit Report after the follow up action. The follow up results will be discussed with the individuals involved

in the original decision for corrective action and any modifications to this corrective action made as agreed.

Therefore, the following documents will be part of the Audit Program Plan:

1. Schedule of audits
2. Completed Audit check Lists
3. Summaries of corrective action agreed upon.

4. Monthly Audie Reports.

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The Audit Check List will be used as the basis for each Audit.

The topics covered by chis List are as follows:

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AUDIT CHECK List

Does the Leader of the scientific unit has the required
basic docunents? (Described in Section V of North West Coast

Project Quality Assurance Progr:

Data Confidence

*) Are samples protected from loss, contamination or change

») Is the calibration of instruments adequate?

©) Is there statistical treatment of data where applicable?

@) Are data self-consistent; if not why?

©) Are the samples preserved carefully where needed?

1) Are the data kept carefully; log-books titled, dated and

signed?

Is the Program progressing according to schedule?

Is time scheduled for writing reports?

Are report deadlines being met?

Review corrective action

---Page Break---

AUDIT cHECK List

(sample)

SCIENTIFIC UNIT

ee

SCIENTIST 1N CHARGE

T+ Boes the leader of the scientific unit has the required basic
?ocuments a5 described in Section V of the North Vest Coast

Project Quality Assurance Progran?

TL. data conti

As Are samples protected from

1. tos
2. Contamination
3. Change

B. Is calibration of instruments adequate?

1. Instruments calibrated at unit, List:

Date Last Calibration

Instrument Calibrated Interval

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2. Instruments requiring outside calibration:

Date Last

Calibration

Instruzent

Interval

?Are calibration dates and intervals on individual
Instruments?

ST there statistical treatment of data where applicable?

Re: Section 5.16.3 of PREPA specifications

>.

Are data self-consistent?

If not, why?

=

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Are samples preserved carefully after study where
necessary?

Are data kept carefully?

1, Log books titted and dated?

2. Entries dated?

3. Entries signee?

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111, 19 the program progressing according to schedule?

VIZ, Is time scheduled for writing reports?

VITI. Are report deadlines being met?

mK,

1, Corrective action and date of agreement.

2. Current status of corrective action.

3 Further recommendation:

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QUALITY ASSURANCE MEMORANDA

(Quality Assurance Memoranda will be distributed

5 it develops to all QA Manual holders, for

incorporation in the same).

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