

CEER-M-002

THE RELEASE OF CADMIUM, CHROMIUM, COPPER, NICKEL AND ZINC
BY SEWAGE SLUDGE AND THE SUBSEQUENT UPTAKE BY MEMBERS
OF A TURTLE GRASS (THALASSIA TESTUDINUM) ECOSYSTEM

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CENTER FOR ENERGY AND ENVIRONMENT RESEARCH

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by

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ABSTRACT

This research was initiated to determine the rates of uptake, by a
Thalassia testudinum ecosystem, of Cd, Cr, Cu, Ni, Pb and Zn which were
leached from sewage sludge by seawater. The experimental design used aerated
flowing seawater (8.4% min!), which passed over 2 0.1 m² bed of sewage

sludge before traversing the model ecosystem. The tanks, both control and experimental, were 9.2 m x 0.9 m x 1.1 m in size with a volume of 3.1×10^3 liters. Each tank contained sand to a depth of 0.5 m for a total volume of 4.2 m³. The experiment ran for 125 days from March, 1975 to July, 1975 and was duplicated from December 1975 to April, 1976.

The largest net uptake of metals occurred in the "fouling organisms" where Cd, Pb, and Zn uptake closely paralleled the net loss of metals from the sewage sludge. *Thalassia Teaves* showed a net uptake for Cr, Pb, Ni, and

In for both experiments. The urchin (*Lytechinus*

ariegatus) a herbivore on

Thalassia leaves also demonstrated a net uptake of Cu, Cr, Pb, Zn, and Ni in

both experiments. The sea cucumber (*Holothuria wexicana*), in both experiments

showed a net uptake of Cr, Cu, Pb, and Zn. Net uptake of metals in mangroves (anizophorae mangle) was limited to the roots. In the first experiment Ni and Zn showed a significant uptake whereas in the second experiment only Cr demonstrated a net uptake. The uptake in mangrove roots appeared to be a direct function of metal concentration in the sediment. The lack of apparent

retal uptake in the sediment, except for Ni, Pb and Zn in the second experiment, was probably due to the sampling technique rather than to a lack of

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uptake of metals by the sediment,

no consistent or significant metal uptake was found for the clam (*Codakia orbicularis*), oyster (*Crassostrea rhizophorae*) or the snail (*Neritina tessellata*).

The lack of sufficient sample mass, for these organisms, probably obscured any net uptake that may have occurred.

The results indicate that the dumping of sewage sludge in coastal tropical waters can lead to the uptake and concentration of toxic trace metals

by members of a turtle grass community.

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SECTION T

INTRODUCTION

The deposition of sewage sludge in the marine environment may adversely affect the resident organisms in three ways: (1) Siltation with resultant death of sessile organisms. (2) Production of an anoxic environment due to the oxidation of the large organic load in the sludge. (3) by the release of toxic materials into water in forms that may be incorporated into the organisms. These materials usually include metals (zinc, cadmium, chromium, nickel, lead and copper), pesticides or organohalogenes and hydrocarbons of petroleum origin. The mass dumping of treated sewage sludge into oceans and shallow inshore areas is occurring in a number of areas. In the New York Bight, an example of an area receiving large quantities of solid waste for almost a century (Carmody et al., 1973). Carmody, Pearce, and Yasso (1973) determined that there was a definite increase in the sediment of Cr, Cu, Pb, Ni and Zn due to the dumping of sewage sludge. The concentration of Cr, Cu, Cd, Pb, Ni, and Zn in sewage sludge is markedly elevated over ambient marine water concentrations (Salotto, B.V. and Farrell, J.B., 1971; Jacobs, S.A., 1973). Therefore, the deposit of this sludge in the marine environment could result in the uptake and concentration within the food web. The uptake of metals by marine organisms and concentration within the food web has been demonstrated numerous times (Jacobs, S.A., 1973; Kerfoot and Jacobs, 1973).

1973; Kerfoot, W.B., 1973; Shuster and Pringle, 1969; Valiela et al, 1974).

The most extensive compilation of metal concentrations in the tissues

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of marine organisms is found in Vinogradov (1953) and Goldberg (1965). However, the results were usually based on very small sample sizes so it is very difficult to assess the background levels of metals in marine organisms.

Eisler (1973) compiled an annotated bibliography on biological effects of metals on aquatic organisms. Montgomery et al. (1976) found that in order

to obtain a valid sample size for trace metal determination in the thread

fin herring (*Opisthonema oglinum* (Le Sueur)) a pooled sample size of from 50 to 124 fish was necessary in order to detect a 15 to 25 percent difference

between two populations. Other researchers have shown the effect of various metals on the marine food web or individual organisms both in laboratory

studies and field experiments (Phelps, O.K. et al., 1975; Nair, K.V. et al.

1973; Eisler, R. et al., 1975; Eisler, R., 1975; Gardner, G.R. and G.

LaRoche, 1973; Jackim, E., 1973; Frazier, J.M., 1976; Ferrell, R.E. et al.,

1973; Huggett, R.J., 1975; Hannan, P.J. and C. Patouillet, 1972). Schroeder

(1975) demonstrated, using radioactive isotopes, that *Thalassia testudinum*

would concentrate cobalt and manganese in the leaves rather than the root structure (excluding rhizomes). The incorporation of cations in *Thalassia testudinum* was primarily accomplished through the leaves (Schroeder, 1975). The use of sewage sludge to amend agricultural soils has been extensively studied (Street, J.J. et al., 1977; Silveira, O.J. and L.E. Sormers, 1977

Turner, R.I

. et al., 1976). However, very little has been published regarding the rates of release of potentially toxic trace metals into complex tropical ecosystems. The necessity for this type of controlled field experiments was explained by Menzel (1977) for the CEPEX experiment. Inherently, it is impossible to duplicate exactly the marine ecosystem due to its spatial and temporal dimensions. Also, the isolation of organisms from their surrounding environment produces results that are ambiguous. Therefore, it is

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unitarily that any 'artificial' system can duplicate conditions in the natural

environment.

It may not be necessary to duplicate in a laboratory the real system, but rather to produce a system that can duplicate itself (Menzel, D.W., 1977), while at the same time attempting to duplicate the gross physical marine parameters, such as light, suspended sediment load, salinity and temperature. In order to do this, a flow-through seawater system with replicate experiments in different seasons is mandatory:

Our objectives in this research were to use a flow-through system of sufficient size and complexity so as to nearly duplicate a tropical marine ecosystem and allow repeatability over varying seasons. This system would allow us to determine the rates of uptake, by a tropical marine community, of Cu, Cd, Cr, Zn, Ni, and Pb which were leached from sewage sludge by seawater. This community was comprised of turtle grass (*Thalassia testudinum*) with its associated sedimental infauna, spiny sea urchin (*Lytechinus variegatus*), sea cucumber (*Volothures mexicana*), attached "fouling organisms", plankton, and the red mangrove (*Rhizophora mangle*).

This ecosystem was chosen for three reasons: (1) the system comprised a tightly bound community with strong interaction between the members, (2) the important role of the seagrass/mangrove ecosystem in the overall

tropical near shore environment (Jones, J.A., 1968; Odum, H.

< et al, 19595

Golley, F. et al., 1962), (3) the possibility of the active concentration of these trace metals in the higher trophic levels with passage of the materials through the food webs.

The results from this research will aid in forming a basis for determin

ing the potential effects of leached toxicants from sewage sludge on a tropical marine ecosystem. These problems are especially critical in Puerto Rico.

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The population density on the island is nearly 2000 people per square kilometer about ten times that of the continental United States. Only a few towns and cities have sewage treatment facilities and most of the sewage is dumped, minimally treated, into the rivers and bays.

With the increased awareness of the potential detrimental effects of raw sewage on the health of the inhabitants and the marine ecosystem, the construction of sewage plants will increase. This increase of modern plants will produce large amounts of sewage sludge. The disposal of this sludge will pose an especially critical problem in the densely populated, mountainous island of Puerto Rico as the available land for sludge disposal is very

limited and disposal poses its own set of ecological problems at least as

serious as marine disposal of sludge. Therefore, the judicious selection of ocean dump sites for sludge, if this alternative is chosen, will require estimates on the leaching rates of toxic substances from sewage sludge by seawater. These substances can enter the food web and become concentrated relative to the concentration in the water column. The toxicity to marine organisms of Cd, Pb, Cr, Ni, Zn and Cu has been demonstrated (Eisler, 1971; Eisler, R. et al., 1972; Frazier, J.M., 1976; Gardner, G. and LaRoche 1973; Ikuta, K., 1968). It is also possible that some of the traceable substances could promote growth in portions of the ecosystem i.e., vitamins, chelators, organic growth substances (Valentine, J.R., 1957). However, the potential possible benefits of sewage sludge leachate will not be examined in this research.

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SECTION 2

METHODS

The seawater system and tanks were located near Branadero Bay, approximately 8 km from Mayaguez, Puerto Rico (Fig. 1). Two tanks (9.2m x 0.9m x 1.1 m) were constructed from 1.9 cm exterior grade plywood with 5 cm x 10 cm external framing (see Fig. 2). These tanks were sealed with two coats of

Fibreglass epoxy resin and leached in running seawater for 2 weeks. The tanks were located in an open structure. The roof of the structure was fabricated of galvanized corrugated roofing. The underside of the roof was covered with plastic sheeting to prevent zinc contamination.

Each tank was filled with 4.2 m³ of calcareous beach sand (7x CaCO₃) from Aguada, Puerto Rico. The mean depth of sand was 0.5 m with an additional 0.1 m of sand over the last 3 m of tank. The tanks were then filled with seawater to a mean depth of 0.5 m and flushed for an additional week.

The seawater for the system was pumped from the dock of the Center for Energy and Environment Research at a depth of 3 meters. The pump was a Sears Dynaglas 1/2 hp (230 volt). A plastic minnow trap was used as a coarse pre-filter for a Filterchem combination filter and foot valve at the pump inlet. ANI piping used was grey schedule 40 P.V.C. The water traversed 73 m to a charcoal, sand and gravel filter (0.5 x 0.5 x 1.0 m), then to a 1000 liter settling tank. The settling tank was constructed of 1.9 cm thick plywood

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Lined with two coats of fibreglass and epoxy resin. The water was then gravity fed to the two seawater tanks with the flow split using P.V.C. 1" Joints. The flowrate in each tank was monitored and maintained at 8.2 to 8.7 l min⁻¹ by PVC ballvalves. The total water volume was 3.1 x 10³ litres. The turnover time was 5.9 hrs per tank. The inflowing water was diverted

below the surface by submerging the inflow end using a perforated inflow pipe (Fig. 2). The seawater was continuously aerated with an air pump and air stones.

Sewage Sludge

Activated sludge was shipped from New York City in 208 L polyethylene lined drums. The sludge was allowed to settle and the overlying liquid decanted. In general, sludge is 5x solids. The black, "oily" sludge slurry remaining was mixed in a large fibreglass tank. The sludge was added to 2 depth of 7.6 m in an area 1 m wide by 2.3 m long (0.1 m) at the incurrent end

of the Experimental tank which had previously been sealed off from the rest of the tank. A PVC frame with small mesh screen attached was placed over the sludge to prevent direct contamination of the organisms. A duplicate screen was placed in the control tank. After 24 hrs, when the sludge had settled, the seal was removed and Experiment I was begun. At days 1, 5, 25, 50, 85 (Experiment II only) and 125, after the sludge was added to the Experimental tank, samples of organisms, sediment, sludge (day 1, 125 EXPI) and water were taken from both tanks. A presample was taken prior to addition of any sludge, in this study it is referred to as day 0.

Sampling

Organisms

The initial stocking quantity or density for the biological organisms, used in both Experiment I and II, are shown in Table 1.

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Table 1. Physical dimensions of organisms with number of total

organisms per tank and number of organisms

supplied per tank per day.

Experiment 1

Experimental Tank ?control Tank

Low emerald a/Sanctet | L__ WH detected #/Serple

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Experinental Tank Controt

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sive ?63 S38 H

Ses Cucumbers i 2

si ?

seats.

oysters

fvsraite ust tien on day 125 control 2s none Tet

supper values are for gays 0 through 50, Experiment 1

over values for day 125. savers refer to naer of

frgsncass in each pooled sample

The organisms, except for the clans, were all collected using manual techniques at Pt. Osteones, Puerto Rico. The organisms were allowed to ?equilibrate for 2 weeks in the flowing seawater system. The turtle grass

was replanted manually at 400 plants m², after soaking in a 10x NAPDH sol~

ution (Kelly et al., 1971). The clams were collected at Pt. Viento on the Southeast coast of Puerto Rico.

Mangroves in Experiment 1 (230 seedlings) were planted as ungerminated seedlings and as germinated seedlings in Experiment I (120 seedlings).

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tysters in Experiment I, were placed in tanks and suspended in one plastic mesh bag per tank. In Experiment II the oysters were placed 13 per bag, for a total of 10 bags per tank. The location of the bags was randomized except no bag was hung directly over the sewage sludge deposit.

Table 2 is a species, common name list of plants and animals used for

this study.

TABLE 2

Species

Clam *Arca*

Oyster *Crassostrea rhizophorae*

snail *Nerita tessplate*

Sea cucumber *Holothurea* sp. (predominantly #. *mexicana*)

Urchin *Ytechinus variegatus*

Turtle grass *Thalassia testudinum*

Red mangrove *Rhizophorae mangle*

On

After equilibration, organisms were sampled at the prescribed inter-

vals, the samples were removed, physical measurements made, placed in double bags and frozen. Simultaneously, samples were collected for hydrocarbon analyses. The samples for hydrocarbon analyses were placed in glass bottles with aluminum foil seals, capped and frozen. The bottles were previously rinsed with 2N HCL, deionized water, CCxq, and air dried. The physical dimensions for the animals sampled and number of animals sampled per day are shown in Table 1.

overlying Water

Water samples were taken, for Experiment I and IT, prior to biological

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samples. There were three types of water samples taken: pore water for anal-

ysis of soluble trace metals and hydrocarbons, overlying water for soluble and particulate trace metals and hydrocarbons, and dissolved reactive phosphate (DRP). The DRP was sampled at weekly intervals. The trace metals and hydrocarbons in water and pore water were sampled at the same intervals as the organisms. Pore water samples were not taken in Experiment 11

Pore Waters

Samples were taken using one inch PVC pipes sealed at one end with 0.8 mm holes drilled in the lower 38 cm of the pipe. The pipes were placed 46 cm from the sides at 1.2, 3.5, 5.8, and 8.1 m from the intake end of the tanks. Two liter water samples were withdrawn using 6.4 mm Tygon® tubing and 25 Tbs of vacuum pressure, from a laboratory vacuum pump.

Sediment,

Sediment was collected using a 2.5 cm diameter x 25 cm PVC core. The sediment core was homogenized and ground after drying (105°C) and a 1g sample taken for trace metals. After day 25 in Experiment I and throughout Experiment II, 25g sample was taken for trace metals.

Dissolved and particulate trace metals

All water samples were placed in 2.5 L glass bottles (which formerly contained conc. HNO₃ or HCl) which were equilibrated in reagent grade seawater for 12 hrs prior to sampling (4 & samples for Experiment 11). The

sample bottles were drained, rinsed with the sample, filled, then filtered within 4 hrs. The filtering was accomplished using an all plastic filter system containing a .4 μ m pore size, 142 mm diameter, Getnan® filters. The filters were previously washed in 6 NHC and rinsed with copious amounts of R.O./D.I. water. The fraction retained on the filter was referred to as the ?particulate? portion. The filtrate was the ?soluble? fraction.

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Physical and Chemical Analyses

General

ANI reagents used are reagent ACS grade. AIT glassware was cleaned with detergent, rinsed with water from a Millipore * »Mi115-9" system. AN? water referred to in this paper is R.O./D.I. water from the Millipore R ?Mi111-Q" system. This refers to the technique of reverse osmosis followed by ion-exchange. The final cleaning for glassware used to digest samples for trace metal analysis involved refluxing the covered beakers with concentrated HNO₃ followed by rinsing with R.O./D.I. water. ANI other equipment was washed with detergent, rinsed with R.O./D.I. water, followed with 6N HCl and rinsed with R.O./D.I. water. Only non-metallic instruments and materials were used and all equipment, after cleaning, was stored covered with Parafilm R or plastic bags.

Oxygen, temperature, and salinity

A Yellow Springs International YSI - 57 oxygen/temperature meter was used. The oxygen output was both salinity and temperature compensated, The oxygen calibration was checked using the Winkler titration technique (Steckland and Parsons, 1965).

Dissolved reactive phosphate

(One hundred ml samples were collected and immediately filtered through a Millipore Swin-Loc R adaptor containing an acid washed (6N HCL) 27 mm Millipore, 0.45 µm pore size Filter using an acid washed 100 ml. polypropylene syringe. The sample was frozen and Total DRP was determined using the single solution technique (Strickland and Parsons, 1965) with a 10 cm cell in a Beckman DU spectrophotometer. A small number of samples were run on

Technicon autoanalyzer using standard Technicon procedures (Zimmermann, C. et al., 197).

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Particulate trace metals,

The 142 µm diameter filters were placed in Pyrex reaction beakers, 6 ml of concentrated HNO₃: HCL (3:1) were added and the beaker covered with a ribbed watch glass. The solution was heated to a slow boil (less than 95°C), when the filters dissolved, the liquid was brought to a rapid boil and taken to

near dryness. Then, 4 ml concentrated HNO₃ were added, followed by dropwise addition of 1 ml of H₂O₂ (30% by volume). The sample was then rebottled to near dryness and 3 drops of concentrated H₂SO₄ were added. If the sample turned black more HgO was added. The samples were dried until SO₃ fumes disappeared. The sample was then brought to 10 ml volume with R.O./D.I. water.

Soluble Trace Metals

After 2.56 of sample was filtered for determination of particulate trace metals (41 in Experiment I1), the effluent was run through two 1.2 cm diameter x 10 cm glass columns. The flow rate was maintained at 1 ml min⁻¹. The columns were in series and the first column contained 10 ml of Chelex-100 in the ammonium form (20 ml in Experiment I1) with 20 ml of Anberlite XAD-2 resin (Rohm and Haas) in the second column. The Chelex resin was purified by 2 bed volumes (BV) of INHCE (isothermally distilled) followed by 5 BV of D.I./R.O. water and then 2 BV TN NH₄OH (isothermally distilled) with a final wash of D.I./R.O. water. The Anberlite XAD-2 resin was cleaned by washing for 10 minutes with 0.1% D.I./D.O. water. The water was decanted and the procedure repeated 5 times. The resin was washed four times with anhydrous methanol followed by four washes with 0.1% D.I./D.O. water. The Chelex resin was left covered with water prior to use. The Anberlite was stored under methanol until just prior to use.

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In Experiment I and IT, the metals were extracted from the Chelex-100[®] resin after the resin column was first washed with 100 ml of water. The trace metals, in EXP I, were eluted with 40 ml of boiling concentrated HCL. The resin was rinsed with 20 ml of water and this water was combined with the acid eluate. The eluate was bottled down (95°C) to 1 ml and then brought to 10 ml volume with water. This was referred to as the soluble inorganically complexed fraction. In Experiment II, the Chelex 100 was eluted batchwise using 6 ml concentrated HNO₃ per 10 ml resin (Davey, E.W. and A.E. Soper, 1975) rather than eluting with concentrated HCL. The Chelex resin was filtered and the acid eluate dried to 1 ml followed by the addition of 6 ml 2N NCR (isothermally distilled) and dried down to 1 ml. This was repeated and the final product was diluted to 10 ml with distilled water. The Amberlite[®] resin was first washed with 100 ml water then the resin was placed in an open glass Petri dish, rinsed with methanol, and the resin ignited. The ignited resin was placed in a muffle oven and the temperature raised in 100°C increments until 400°C. The resin was maintained at 400°C for 1 additional hour then the temperature was raised to 450°C for 24 hours. The cooled resin was dissolved in 5 ml INHCL, then boiled down to 1 ml and brought to the desired final volume. This fraction was referred to as the soluble, organically complexed trace metal (Montgomery, J.R. and J.E. Echevarria, 1975). The trace metals were determined by atomic absorption spectrophotometry (A.A.S.) on a Perkin-Elmer Model 303 A.A.S. with a

deuterium background corrector. The output was recorded using a Model 56 Perkin-Elmer recorder and DCR-1 digital readout. The flow chart for sample collection and processing 1s shown in Fig. 3.

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Analysis of National Bureau of Standards (NBS) bovine liver and orchard leaves were performed along with the regular samples. One gran samples were used, when available, for all standards and samples. The flow chart for sample collection 4s shown in Figure 3.

The number of organisms sampled per day per experiment are shown in Table 1. The samples were defrosted after collection and dissected into the following tissues using Pyrex glass shards and plastic gloves.

Urchins = shell and internal tissues.

Sea cucumbers - body, muscle bands and internal tissues. In Experiment TL, muscle bands were combined with internal tissues

Snails, clams, oysters - shell and internal organs.

Fouling organisms - total mass scraped from glass collecting plates

(23 x 30 cm). In Experiment 1, all plates were sampled at each sampling

tine, In Experiment 11, cumulative results were obtained by sampling plates which had remained in the tanks for 25,50,85,125 days after addition of sludge.

Thalass'

= Roots and leaves. The roots and rhizomes were combined and are referred to as roots.

Mangroves - The lower 3.8 cm of the hypocotyl not including the roots.

This is referred to in this paper as the bottom, Growing stem includes a) of the primary meristematic tissue from the hypocotyl excluding the leaves and the roots/rhizomes.

After dissection into these various parts, the wet weight was obtained.

The samples were dried in covered, cleaned Pyrex beakers at 105°C. The dried

sample was weighed, then homogenized, using a ceramic mortar and pestle.

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TABLE 3

Results in ug g⁻¹ dry weight for determination of metals in NBS bovine liver and orchard leaf standards. The results for Experiment I and II are shown with statistics and our computed minimum detectable concentration (MDC).

EXPERIMENT 1

Bovine Liver

ca cu cr Pb In Ni

oc 0.3 8.9 3.4 0.5 28 09

R 0.56 7 2.5 132 -

so 0.26 20.2 98 a7

N 9 10 10 i

EXPERIMENT II

co cu cr Pb an Ni

x 0.55 187.8 WC OTL.

so 0.45 15.50 17.59 0.69

N 49 4a 53 48

Nes 0.27 4.04 193810 = 0.383 0.08 1304 10

values

EXPERIMENT I

Orchard Leaves

i 0c 2.300 26 44.8 26.1 3.9

3.9 (0632.82 54 0.6

n 6 6 n 3

EXPERIMENT 11

Orchard Leaves

x 0.66 u - 5.0 25.6 4.0

so 0.34 15 3.6 7.64 13

N 7. 20 23 26 24

fos COMER EET (ES) BET ET TST Oe

Va

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The ground sample, not used for analysis, was stored in plastic, screw cap potties. The dried samples were placed in cleaned Pyrex beakers, at least 1g where possible, and were digested using the following procedure.

Twenty-five ml of a concentrated HNO₃:HClO₄ mixture (3:1) was added to the dried, weighed sample in the covered clean beaker. The sample was slowly boiled and refluxed in the covered beaker at 90 to 95°C until 1 ml of solution remained. The sample was allowed to cool then 30 ml of 30% H₂O₂ was added slowly and reheated to 90-95°C until 1 ml of the solution remained.

A further 10 ml of HNO₃:HClO₄ mixture was added and reheated. The mixture was allowed to cool and 10 ml of 30% H₂O₂ was added. This was allowed to boil again (90-95°C). The 10 ml additions were repeated, if necessary, a maximum of three times. Six ml of 2N HClO₄ were added, boiled at 90-95°C, until 3 ml were left. The sample was allowed to cool then centrifuged in a clean 50 ml

plastic capped centrifuged tubes at 1700 rpm for 20 minutes. The liquid was carefully decanted and brought to 10.0 ml with R.O./D.I. water. This sample was placed in clean, 25 ml polyethylene vials with caps.

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SECTION 3

RESULTS

The raw data, expressed as ug metal/g dry weight for all samples, are found in Appendix A. The computed net uptake for each metal is located in Appendix B and is expressed as mean experimental results (9/9 dry weight) minus the mean control results for each sample, with their respective standard deviations, and degree of freedom. The t value for the null hypothesis, $H_0: \mu_M - \mu_C = 0$ ($P < 0.05$) where μ_M is experimental and μ_C is control is, also shown in Appendix 8. The analyses for the NES biological standards are shown in Table 3. The results for all fractions of dissolved and particulate trace metals in the water for both control and experimental tanks (Experiment 11) are in Appendix C. The wet/dry weight ratios with statistics for all the samples in Experiment I and 11 are in Appendix D.

The experiments were replicated, Experiment I began on 11 March, 1975 and ended 15 July 1975; Experiment 11 began on December 12, 1975 and ended April 26, 1976. The experimental tank refers to the tank with the sludge

added and the control tank was identical, in experimental design, to the experimental tank except no sludge was added.

Temperature, salinity, and oxygen

In Experiment I the temperature ranged from 21 to 26°C with a salinity range from 32 to 34 ‰. In Experiment II the temperature ranged from 21 to 26.5 with a salinity range from 28 to 35 ‰ (¥, 30 ‰, \$.0. 4.2 ‰. N= 18).

15

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Dissolved reactive phosphate (DRP)

The results are shown in Figs. 4 and 5 for control, Experiment 1 and 11 and experimental, Experiment I and 11, respectively. The plotted points are the mean values for a five day period.

Fouling Organisms

In Experiment 1. Cd, Cu, Pb, and Zn showed a significant net uptake after 125 days (Figs. 6, 8, 9, 11), whereas in Experiment II, Pb (Fig. 10) showed a steady uptake to the end of the sampling period (85 days). For Experiment II, Cd and Ni demonstrated a steady uptake until day 50 (Figs. 7 and 13) then a decline in the net uptake. Zinc and chromium indicated a net

uptake after day 50 (Figs. 12 and 15) and Cu showed little significant uptake for the sampling period (Appendix B).

The initial values for trace metals in sewage sludge are shown in Table 4.

A rapid decline was noted in the concentration of Cd, Cr, Cu, Ni, and Pb in the sewage sludge for Experiment I (Figs. 6, 15, 8, 14, 11). The Pb results are inconclusive for both experiments (Fig. 9 and Appendix B) due to the loss of lead as the easily volatilized $PbSO_4$ during the digestion of the sludge. Because of the shorter sampling interval for sludge in Experiment 11, a two-stage decline for Cd, Ni, and Zn was shown (Figs. 7, 13, 12). The first decrease in metal concentration was nearly instantaneous with a further, smaller decrease after day 50.

Thalassia

Leaves

The results of EXP I were biased due to the die-off of the plants by

---Page Break---

day 50. This was probably due to insufficient light. The plants survived for the full 125 day period in Experiment II after the Tight Tevel was increased.

Copper, Crs Pb, Nf and Zn showed rapid increases in net uptake in Experiment I (Figs. 16,18,20,22,24). In Experiment II the same overall trend of increasing net uptake was shown for Cu, Cr, Pb, and Cd for Experiment 11 (Figs. 17,19,21,26). Mekel and zinc, in Experiment 11 did not show @ significant uptake up to day 50 but the net uptake was significant at day 85 and 125 (Figs. 23, 25).

Roots

The only significant net uptake of trace metals for Thalassia roots and rhizomes in Experiment I was for nickel, Cr, Cu, and Pb (Figs. 27.29.31, 33).

The repeat experiment also showed a significant net uptake for Cr and In (Figs. 30, 36) with Pb, Ni, and Cu showing peaks of significant net uptake values at day 5 and/or 25 days (Figs. 34,28,32). Cadmium uptake was significant only at day 25 (Fig. 37)

Urchin, internal organs

To facilitate later discussion, the results of the net uptake for urchins

are shown with the results for Thalassia leaves.

Copper, Cr and Pb show a steady and rapid increase in net uptake for EXP I and II (Figs. 16,17,18,19,20,21). The net uptake for Cd was not significant in Experiment I (Appendix 8), however in Experiment II (Fig. 26) the net uptake increased after day 50. The results for nickel and Zn uptake are very similar in Experiment I and II (Figs. 22,23,24,25,26). Zinc shows

a steady increase with a decrease, in net uptake, at day 50 followed by

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rapid increase in both Experiment I and II

Sea cucumber, internal organs

In both Experiment I and II, Cr, Cu, Pb and Zn showed very similar net uptakes over the 125 day period (Figs. 38,39,40,41,42,43). Ni showed a net uptake spike at day 5 in Experiment I, and in Experiment II a spike showing uptake of nickel occurred at day 5, 50 and a smaller spike at day 125 (Figs. 38, 41).

Mangrove Roots

Nickel, in Experiment I, shows an initial decrease in net uptake before day 50 with a gradual increase to day 125 (Fig. 27). The opposite occurs in Experiment I1 where the net uptake increases rapidly up to day 5 (Appendix 8). The net uptake for Cr was not significant for Experiment 1 (Appendix 8), but indicated a linear net uptake from day 50 to day 125 for Experiment 11 (Fig. 30). Lead uptake in Experiment I and 11 (Figs. 33 and 34) exhibits similar trends. However, Pb. uptake is much higher in Experiment 1 (Fig. 33). Zinc, in Experiment 1,

1s0 demonstrated @ rapid linear

uptake curve (Fig. 35) which was very similar to the uptake in Experiment IT (Fig. 36). However, the net uptake was greater for 2n in Experiment 1. The net uptake for Cu was not significant in Experiment 1 (Appendix 8), but a

peared to show a gradual uptake after day 50 followed by a decline to original levels (Fig. 32).

other

No stontficant net uptake was shown for the oysters, clams, mangrove body, stem or leaves, snaTs, holothurian body and muscle parts or sediment

for Experiment I or II. The results shown are in Appendix 8. The significant results for net uptake of metals are shown in Figs. 6-43 and are indicated by

open rectangles. Dark rectangles indicate non-significant results.

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TABLE 4.

Concentration of Cd, Cr, Cu, Ni, Pb and Zn in $\mu\text{g/g}$

dry weight, in sewage sludge used in this experiment.

The results are for Experiment 1 and II.

Results for the same metals as determined by

Salotto and Farrel) (1971) for activated sludge

are included.

Experiment

Cd Cr Cu Pb Ni Zn

x 109 2120. 2860. 603192926

SD 6.70 115. 124.0 5511.3 388.4

Experiment 11

C4 Gr cu Poe Wi Fy

% 166. 36672445. 21.0 376, 2508

SD 85.5 338.5 266.4 © 3.29 12.7 122.1

Salotto and Farrel] (1971)

ctr cu POF ONE Zn

350 4330 11001500 3803300

Lead values are low due to digestion technique:

rt

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Addition of sewage

Collection of sediment/ Collection of water

Sludge/organisms (0,1, samples for trace

5,25,50,85%, 125 days) metal and hydro-

carbon analysis,

{ES

sample taken*

Frozen in plastic bags for analysis Filtered

of hydrocarbons

Defrost Frozen

sample for* hydro- R

Dissection carbon analysis Anberlite XAD-2

resin columns and

Sample pooted by tissue Extracted with CCL, Chelex-100 * resin

??J Extract stored in * column

?Subsample* freezer |.

mercury

analysis Elution, dilution

wet weight |

o Analysis, atomic

Dry Toss absorption

Dry weight

Grind by

mortar/pestle

Digest/dtution

1

Analysis,

atomic absorption

samples were collected at 0,1,5,25,50,85** and 125 days after addition of

sewage sludge.

?Samples for mercury, and hydrocarbons in both water, sediment, sludge and organisms were stored frozen for future analyses.

an additional sampling was made for Experiment II, Day 0 refers to the sample taken before addition of sewage sludge.

FIGURE 3. Flow chart of trace metal and hydrocarbon sampling from collection processing into final sample form.

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SECTION 4

discussion

Oysters and clams have been extensively studied regarding trace metal uptake because of their commercial significance, hardiness and feeding method. Bivalves, in general, show uptake and concentration of most trace metals (Kerfoot, W.B., 1973; Kerfoot, W.B. and S.A. Jacobs, 1973 a and b; Shuster, C.N. and B.H. Pringle, 1969; Huggett, R.J. et al., 1975 and Frazier, J.N., 1976). However, oysters and clams in our research showed no consistent nor significant net uptake of metal except possibly zinc and copper. In Experiment I

all the oysters were placed in a single net bag.

This procedure was changed to ten net bags per tank in Experiment IT. This drastically reduced the crowding and increased the survival of the oysters beyond day 50 in Experiment IT. The wet weight of the clam and oyster internal organs was too small to allow an adequate sample size for the trace metal analyses. This same problem was encountered in the analyses for the snails, Holothurian muscle parts in Experiment 1, and zooplankton samples for both Experiment I and 11.

The results for lead in the sewage sludge were very low compared to other investigators (Table 4) and this was due to the addition of H_2SO_4 to aid in the digestion of the sludge organic matter. When the sample was heated, the lead was lost as volatile PbSO_4 . The H_2SO_4 was only used for the digestion of the sludge.

The results for the analyses on the NBS bovine liver standards (Table 3)

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are in good agreement for Cu and Zn and high for Cd. Our results for Cr and Pb in bovine liver were below our detection limits. The results for NBS orchard leaves were in good agreement for Cu, Pb and Zn. Cadmium, in NBS orchard leaves, was too high as was nickel. The coefficient of varia

tion (CV) for Ni was lower than the CV for Cd. The results for Cr, in orchard leaves, were in good agreement with the non-certified value for Cr published by NBS.

Our results for the concentration of trace metals in water for Experiment I were at or below the detection limits for nearly all metals and were therefore not shown. This was due to the small sample size. When we used a larger sample in Experiment II, the results for most of the metals were within our detection limits. However, no significant difference was noted between metal concentrations in the experimental tank and those in the control tank. Obviously, metals were released from the sludge, as shown by the results in Figs. 6 through 15 and, also, by the significant net uptake of metals by *Thalassia* leaves and roots, urchin and sea cucumber internal parts, and fouling organisms. This net uptake in organisms was hard to explain when no net change was seen in the water of the experimental tank. The problem in interpretation of the water results was due to the "point sampling" of the water versus the "integrated" sampling for the organisms, sediment, and sludge. The "point" concentration of the water was very low at any one time (Appendix C). These trace concentrations, coupled with the high blanks and matrix problems in trace metal analyses of seawater by flame atomic absorption spectrophotometry, produced results with very high within-sample variance. This high variance effectively obscured any differences between the experimental and control tanks in Experiment II. Schuster and Pringle

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(1969) demonstrated that the concentration of metal in the water, at any one time, may not be the best indicator of the total contamination by metals.

The net uptake of metal by "fouling organisms" was clearly shown in both Experiment I and II. This net uptake was replicated in Experiment I and II for Cd, Pb, and Zn. The uptake of copper by fouling organisms was only significant in Experiment I and for Ni and Cr in Experiment II. The fact that Ni and Cr didn't show a net uptake in Experiment I was probably due to a change in sample technique for fouling organisms initiated in Experiment II. In Experiment I, when a settling plate was sampled for fouling organisms, clean plates were added for the next sampling period, whereas, in Experiment II, a plate which was sampled at day 125 had remained in the system from day 0 to day 125. The fouling organisms sampled in Experiment II are, therefore, integrated samples over the sampling time. The net uptake by the fouling organisms of Cd, Pb, and Zn closely follows the loss of these same metals from the sewage sludge for both Experiment I and II. This same correlation, between fouling organisms and sludge, holds true for Cd and Ni uptake in Experiment I.

The net uptake by *Thalassia* leaves and urchin internal organs closely parallel each other for Cu, Cr, Pb, Ni and Zn in both Experiment I and II. Significant cadmium uptake by *Thalassia* leaves and urchin internal organs was seen only in Experiment II. Schroeder (1975), using radioactive ^{57}Co , ^{60}Co , ^{64}Ni and ^{65}Zn in seawater, showed that, in *Thalassia testudinum*, the

rhizomes had the highest uptake followed by leaves then roots. In both Experiments our results for *Thalassia*, show significant net uptake for Cu, Cr, Ni and Pb in roots. The net uptake was higher in Experiment I for Pb

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land Cu and higher in Experiment 11 for Ni and Cr. Cadmium and zinc had a net uptake in *Thalassia* roots only in Experiment 11, This uptake of metals by the urchin and the close correlation between the urchin's uptake and the uptake in *Thalassia* leaves was not surprising as the urchins were voracious foragers on *Thalassia* leaves and epiphytes. This was also shown in the field by Camp et al., (1973). The net uptake of some metals in leaves was closely coupled to the uptake in the roots of *Thalassia*. The results for Cd, Ni, and Cr in Experiment II indicate a translocation of these two metals from roots to leaves. The translocation of metal was also indicated for Zn in Experiment II from day 85, where the net uptake decreases in the roots with an increase in the leaves and vice versa at day 125. In general, the maximum net uptake in *Thalassia* leaves was slightly greater than the uptake in roots. This is in apparent contradiction of Schroeder's results (1975) where roots were found to have the greatest uptake. However, our root sample consisted of roots plus rhizomes and, as rhizomes showed the highest net uptake in Schroeder's paper (1975), the contradiction between our results and Schroeder's is cleared up.

The interpretation of the significant net uptake for mangrove roots indicates that only Ni and Zn in Experiment I show a near linear increase over time, In Experiment 11, only Cr showed a linear response over time, whereas, Zn and Cu Showed an increase in net uptake followed by attainment, of apparent equilibrium then Toss of the metal. Lead shows no net uptake until day 85, These results appear to be loosely correlated with the fact that the sediment never showed a net uptake until at day 8 in Experiment I] when Ni, Pb and Zn begin to show a slight net uptake (Appendix 6) in the sediment. Since no other part of the mangroves sampled indicated a signifi-

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cant uptake of metal (Appendix 8), then the only source for the metals in the mangrove roots must be the sediments. If that is so, then only Ni, Pb and Zn should show net uptake in mangrove roots and then only after day 85 in Experiment II, This lack of significant uptake of metals by the sediments could be due to the pooling of whole sediment samples from a 2.5 cm diam. x 25 cm core. If the gradient of trace metal concentration in the sediment was very steep, then the pooling of the core could have diluted the higher surface metal concentrations with the much lower metal concentrations in the bottom of the core.

There was similarities in net uptake by Holothurian guts for Experiments I and 11. Although there are similar patterns of uptake in the Holothurian

gut, the maximum values for net uptake were always higher in Experiment 1 for Ni, Cu, Pb and Zn. Chromium showed a higher maximum net uptake in Experiment II for Holothurian guts. In Experiment I, the sewage sludge showed a two stage release. The first stage occurred at day 1 followed by a slow constant release until day 50 when another rapid release occurred. A slow but steady release would more likely result in a higher maximum uptake, whereas, a two stage release, as shown by the sewage sludge in Experiment II would, due to the estimated 6 hour residence time for the water in the tanks and 8.5 L min⁻¹ flow rate, produce a lower maximum net uptake. The data for the net loss of metals from the sewage sludge was only available for two sampling times in Experiment I, so it is difficult to determine the true instantaneous net loss of metals from the sludge.

The original objectives for this research, as stated in the Introduction, have been obtained. A definite net uptake was found in members of a tropical marine community. These results were replicated at two different seasons

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and the results, considering the complexity of the system, agree quite well. Originally, the fish, *Diodon helacanthus* (puffer) was to be added to the system. However, after collecting over 360 specimens we found that they were too sensitive to environmental factors other than sewage sludge and were easily killed. Therefore, fish were deleted from the ecosystem in

order to expedite the completion of the experiment.

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SECTION 5

CONCLUSIONS AND RECOMMENDATIONS

Net uptake of trace metals by members of a Thalassia

testudinum/

Rhizophorae mangle community was demonstrated in two 125 day replicated experiments over the period from March 1975 to April 1976. The net uptake was especially significant in: *Thalassia testudinum* leaves and roots/

rhizomes, internal organs of the urchin *Lytechinus variegatus*, "fouling

organisms", internal organs of *Holothures* sp., and for some metals in

Rhizophorae mangle roots. In general, the net uptake pathways followed the trophic levels in the food webs. This pathway was especially noticeable

?from the water to the fouling organisms and from the water to the *Thalassia testudinum* Teaves and thence to the urchin herbivore *Lytechinus varieatus*. The net uptake in the *Holothurea* sp. were closely related to the net uptake in the fouling organisms and the trace metal loss rate of the sludge. The food web in this artificial system was dominated by the fouling organisms as would be expected. The uptake of trace metals, leached from the sludge, was always greatest in this complex group.

The results of long term exposure, in a simulated tropical ecosystem, to toxic trace metals more closely approximates the situation in nature than laboratory experiments. In this respect we are in agreement with the statement by Phelps et al. (1975) "Chronic exposures to elevated metal levels through the food web should be used as a more realistic test for possible deleterious effects from such metals as Cr. No artificial system can ever

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reproduce a natural ecosystem. However, it is more important to closely approximate the natural system with a system which can duplicate itself (Wenzel, O.M. 1977). Our results show that chronic exposure to toxic metals can lead to uptake and concentration of these metals in marine organisms. We were able to duplicate our results using this complex simulation of a marine ecosystem.

More detailed work on the net uptake of toxic metals, using this complex

simulated ecosystem, and the roles of the "fouling organisms" and plankton in this system is needed. This must involve integrated sampling of the organisms as well as the water. The advent of more precise methods of direct trace metal determination in seawater using anodic stripping voltammetry and flame atomic absorption spectrophotometry, in lieu of flame AAS, should allow a reasonable sample size for both organisms and water.

The authors would not recommend, based on the results of this research, the dumping of sewage sludge in shallow, tropical marine environments. The

potential exists for rapid concentration of toxic trace metals by members

of the food web high on the trophic scale. These toxic levels could then affect organisms used as food by man.

The next logical step in future research would be to add vertebrate organisms to this system to see if a herbivore, which normally consumes,

turtle grass, would show a net uptake of toxic trace metals.

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Figure 1. Study site location

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Figure 2. Diagram of experimental design, tank configuration, and seawater system

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Addition of sewage

CoNtection of sedinent/

Sludge/organisms (0,1,

5,25,50,85°, 125 days)

sample taken*

Frozen in plastic bags

Defrost,

Dissection

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Sample pooled by tissue

Subsanpie

morcury

analysis

wet weight,

|

Dry 105°C

Dry weight

Grind by

mortar/pestle

Digest/ditution

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Analysis.

atomic absorption

for analysis

of hydrocarbons

sample for* hydro~

carbon analysis

Extracted with CCL,

Extract stored in

freezer

Collection of water

samples for trace

metal and hydro~

carbon analysis,

Filtered

Frozen

os

Anberlite XAD-2

resin columns and

Chelex-100 R

see

Elution, dilution

Analysis, atomic

absorption

Samples were collected at 0,1,5,25,50,85** and 128 days after addition of
Sewage sludge.

?Samples for wercury, and hydrocarbons in both water, sedinent, sludge and
forganisms were stored frozen for future analyses.

?*#An additional sampling was made for Experiment 11, Day 0 refers to the sample taken before addition of sewage sludge.

FIGURE 3. Flow chart of trace metal end hydrocarbon sampling from collection processing into final sample form.

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Figure 4.

ORR. CONTROL Exe. 1

AIG-AT/L

ORP- CONTROL Exe. 2

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crys

Dissolved reactive phosphate concentration Experiment I and II,
control. Data points are mean values for 5 day period,
excluding day 1.

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Figure 5.

eRe,

AIG-RT/L

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a

AIG-AT/L.

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EXPERIMENTAL Exe 1

EXPERIMENTAL Exe. 2

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Dissolved reactive phosphate concentration Experiment I and I,
experimental. Data points are mean values for 5 day period,

excluding day 1.

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<> FOULING ORGRANISMS- EXP. 1

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<o SEWAGE- Exe. |

NERN PPM

Figure 6. Mean net uptake of Cd between experimental and control, fouling organisms and sewage - Experiment I.

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MERN PPM

Figure 7. Mean net uptake of Cé between experimental and control, fouling organisms and sewage - Experiment 1.

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Figure 8.

<U FOULING ORGANISMS- Exe. 1

MERN (E-C> / PPM

<u SewAGe- exe. |

MERN PPM

Hean net uptake of Cu between experinenta} and con
organisms and sewage - Experiment I. eral touting

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Pe FOULING ORGANISMS.

MERN (E-C) /PPH.

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Figure 9. Mean net uptake of Pb between experimental and control, fouling organisms and sewage - Experiment 1

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?pL aunbLy

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MERN (E-C) /PPM

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Figure 11. Mean net uptake of Zn between experimental and control, fouling organisms and sewage - Experiment 1.

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ZN FOULING ORGANISMS? Exe. 2

MERN (E-?) /PPM

ZN SEWERAGE- Exe. 2

MERN PPM

Figure 12. Mean net uptake of Zn between experimental and control, fouling

organisms and sewage - Experiment IT.

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Figure 13.

MERN <E-φ) PPM

Ni Sewee- exe. 2

Mean net uptake of Ni between experimental and control, fouling organisms and sewage ~ Experiment II.

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MERN PH

MERN PPM

Figure 14.

Mean net uptake of Cr and Ni between experimental
and control, sewage ~ Experiment I.

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MEAN CE-C) PPM

NERN PPM

Figure 15. Mean net uptake of Cr between experimental and control,
fouling organisms and sewage - Experiment 11

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Figure 16.

Mean net uptake of Cu between experimental and controls

Thalassia leaves and urchin guts - Experiment 1.

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Figure 17.

CU THALASSIA LERVES- Exe. 2

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cu urchin GUT EXP. 2

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Mean net uptake of Cu between experimental and control ,

Thalassia leaves and urchin guts - Experiment II

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Figure 18.

MERN ϕ E-C) / PPM

eR URCHIN GUT- EXP. 1

> /PPM

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Mean net uptake of Cr between experimental and control,

Thalassia leaves and urchin guts - Experiment 1.

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?R THRLASSIE LERVES- Exe. 2

MERN CEC) PPM

MERN (E-C) PPM

Figure 19. Mean net uptake of Cr between experimental and control,

Thalassia leaves and urchin guts ~ Experiment. 11.

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Figure 20.

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Hean net uptake of Pb between experimental and controls

Thalassia leaves and urchin guts - Experiment 1.

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NERN (E-?) /PPH

Figure 21. Mean net uptake of Pb between experimental and control,
Thalassia leaves and urchin guts ~ Experiment 11.

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Figure 22.

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Ni URCHIN GUT- Exe. 1

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Mean net uptake of Ni between experimental and control,

Thalassia leaves and urchin guts = Experiment: I.

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Figure 23.

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MEAN (E-C) PPM

Mean net uptake of Ni between experimental and control ,

Thalassia leaves and urchin guts = Experiment I]

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EN URCHIN GUTS exe 5

onvs

Figure 24. Mean net uptake of Zn between experimental and control.

?Thalassia leaves and urchin guts - Experiment I.

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2N THALASSIA LEAVES- exe. 2

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?2m URCHIN GUT- Exe. 2

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Figure 25. Mean net uptake of ^{222}Rn between experimental and control.

Thalassia leaves and urchin guts ~ Experiment 11.

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<> THALASSIA LERVES- Exe. 2

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Figure 26. Mean net uptake of Cd between experimental and control,
Thalassia leaves and urchin guts - Experiment 1

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Figure 27. Mean net uptake of Ni between experimental and control,
Thalassia roots and mangrove roots ~ Experiment I.

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MEAN ϕ E-C> /PPM

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NI THALASSIA RODTS? EXP 2

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Figure 28. Net uptake of Ni between experimental and control.

Thalassia roots and mangrove roots - Experiment 11.

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Figure 30. Heap net uptake of Cr between experimental and control,

Thalassia roots and mangrove roots - Experiment {1

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Figure 32.

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Thalassia roots and mangrove roots - Experiment 17

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Figure 34. Mean net uptake of Pb between experimental and control,
Thelassia roots and mangrove roots - Experiment II!

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Figure 36. Mean net uptake of Zn between experimental 200 control.

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HERN <E-C) PPM

Figure 38. Mean net uptake of Cr and Ni between experimental and control, sea cucumber guts - Experiment I

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Figure 39.

o HOLOTHURIAN SUT- Exe. 2

MEAN (E-C) / PPM

MEAN «E-C) / PPM

Mean net uptake of Cd and Cr between experimental and control, sea cucumber guts - Experiment 11.

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Figure 41. Mean net uptake of Cu and Ni between experimental and control, sea cucumber guts ~ Experiment IT.

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Figure 42. Mean net uptake of Pb and Zn between experimental and control, sea cucumber guts ~ Experiment I.

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Figure 43.

Pe HOLOTHURIAN GUT- Exe. 2

MEAN (E-C) /PPM.

MEAN (E-C) /PPH

Mean net uptake of Pb and Zn between experimental and control, sea cucumber guts - Experiment IT.

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Appendix A

Sorted Raw Data Experiment I and 11

There are eleven(11) columns shown which represent the following:

column

1 = Animal type

Clam

Holathurian

Nangrove

T= Thalassia

P = Fouling Organisms

Urchin

S = Sediment

2 - Tissue type

Muscle

internal contents (guts)

Shell

Body/sotton

Leaves

Roots and rhizomes

G = Upper Heristem

= Solids

3. Experiment number ~ 1(1) or 11(2)

4 ~ Sample type - whether from experimental tank (0) or control tank (1)

5 = Represents sample time

0 = presample

1 = 1 day after sewage added

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6 = Concentration (ug/g dry weight) Cd

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Multiple values for the same sample represent replicate samples. The value -999.9 is an indication of a value that could not be detected. All manipulation of the data used one half minimum detectable concentration (MDC) when a value was less than or equal to the MDC. The MOC values are listed in the Results (Table 4).

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APPENDIX 8

Results for net uptake for Experiments I and IT

A Students t test ($P < 0.05$) was eleulated for the null hypothesis:

Ho: $\mu_y = \mu_z$ where μ_y is experimental and μ_z is control for each metal for each combination of Experiment I or II, at each sample time, with $n-2$ degrees of freedom shown (Deg). Also shown are the values of \bar{y} mean-

\bar{z} mean), the individual means and standard deviations.

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[APPENDIX C

Experiment 11, Mater Data

The colum headings are defined as follows:

HC E is data from experimental tank

ch control .

METAL concentration (ug/2) shown are for the defined

metal, either Cd, Cr, Cu, Ni, Pb or Zn

3. MEAN PART values shown represent mean (19/2) concentration
for particulate fraction

SD PART represents the standard deviation of the mean
particulate function

5. MEAN SOC values shown represent mean concentration (19/2)

for the soluble organic fraction (Retained on
Anberlite resin)

6. sp soc represents the standard deviation of the mean
soluble organic fraction

7. MEAN SIC values shown represent the mean concentration
(ug/t) of the soluble inorganic fraction
(Retained on Cheiex-100 ® resin)

8. sp sic represents the standard deviation of the soluble
inorganic fraction

139

---Page Break---

MEAN SOC SD SOC MEAN SIC SD SIC

?SD PART

METAL MEAN PART

cd

ec

?8

cu

Ni

140

---Page Break---

[APPENDIX D

Wet/Ory Ratios

Wet/dry ratios are shown for each sample and tissue type for Experiments
Land 11. These values may be multiplied by the metal concentration (19/9

dry weight) values to obtain concentrations in ug/g wet weight.

441

---Page Break---

Mean Wet/Dry

deviation(\$b)

Clan

Gut

Shel

Sea cucumber

Body

Gut

Muscle

Urchin

Gut

She1t

Snail

Gut.

?She

oysters

Gut

Shev)

Thalassia

Leaves

root/

Rhizomes

"Fouling

organisms"

Senage

Sediment

Mangroves

Bottom of

hypocoty?

Meristem

above

hypocotyl

Leaves

Roots

10

Ratio (3) with n (number of samples) and Standard

fe

for all organisms sampled in Experiment 1 and? {i

10.2

78

5.2

14.8

14

33

3.4

45

5.4

1.90

aa

2.2

7.10

0.00

0.60

0.60

1a

1.0

142

N x 8.0

13 10.7 1.20

12 Va 0°00

12 45 0.40

13 10.6 3.20

Not determined, combined

with out

12 10.7 2.30

13 2.0 0°20

7 3.9 1.0

7 i 0:0

le nes 2.10

13 13 0:20

12 lee 2.90

3 9.9 22

7 5.0 1a

7 8.7 4a

13 14 0.10

7 3.4 0.60

7 4.0 0.40

5.3 1.3

5.6 0.50

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